Ras and p53 expression in non-small cell lung cancer patients: p53 over-expression correlates with a poor prognosis

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Abstract. Expression of the tumor suppressor gene p53 and the ras oncogene were examined in 46 tumor and nodal specimens of non-small cell lung cancer (NSCLC) using the antibodies p53 pAb 240 and ras Y13-259 respectively. p53 expression was elevated in 46% and ras p21 was overexpressed in 85% of the tumor specimens analyzed. Fifteen cases of benign lessions were also assessed for both ras p21 and p53 expression; all were found to have negative staining. p53 over-expression was found to correlate with a poor prognosis in both the tumor specimens (p<0.05) and in the nodal tissues (p<0.005). Ras p21 over-expression was found to be associated with survival (p<0.1) in both the tumor and the nodal specimens. Stage of the disease correlated with survival; similarly both p53 and ras p21 over-expression correlated with stage. No correlations were found with the pathological grade of the tumors nor with a history of smoking or duration of smoking. No K-ras mutations at codon 12 were observed in a further 15 NSCLC specimens analyzed. These results indicate that the p53 gene in particular plays a role in the stages of NSCLC.

Introduction

Lung cancer is the most frequently occurring fatal cancer in the western societies. Even with the best current approaches of early detection less than 10 per cent of all newly diagnosed patients will be cured. Non-small cell lung cancers (NSCLC) account for approximately 75 per cent of all lung cancers (1). Recent advances in molecular biology of lung cancer has provided a genetic framework in which the pathogenesis of the disease may be described (2,3). The evidence to date suggests that changes in dominant oncogenes and in tumor suppressor genes are most likely a

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prerequisite for malignant transformation (4-5). Mutation in the *ras* oncogene family and in the p53 tumor suppressor gene appear to be the most frequent genetic events in lung cancer (6-9). Mutations in the K-*ras* gene have been found by a number of research groups in NSCLC's (10-16) and K-*ras* mutations have been shown to be associated with a history of smoking in adenocarcinomas (6,7,10). The *ras* p21 protein has also been found to be over-expressed at a high frequency in NSCLC compared to SCLC (17,18). Evidence from both NSCLC cell lines and fresh tumor specimens has indicated that the K-*ras* gene is particularly implicated in the development of adenocarcinomas of the lung.

Approximately half of the adult cancers, including lung, breast, colon, esophagus, and skin cancers contain p53 mutations (19) and aberrant p53 expression is now considered to be one of the most common genetic features in a wide range of human cancers (20). p53 has recently been shown to have a biochemical role as a specific transcription factor and to have a biological role as a G1 checkpoint control for DNA damage (21-23).

A number of authors have described p53 mutations in lung carcinomas (8,9,24-26) and also over-expression of the p53 proteins (27-29). However, there is little information to date concerning the clinical outcome of lung cancer patients with regard to mutations or over-expression of the p53 tumor suppressor gene in both tumor and lymph node metastasis specimens.

In the current study we have evaluated both *ras* p21 and p53 expression in 46 NSCLC and analyzed this data with a range of clinico-pathological parameters, smoking history and survival.

Materials and methods

Patients and Pathology. Forty six specimens of surgically treated lung cancer patients and fifteen specimens of benign lung lesions from patients treated at Metaxa Anticancer Hospital in Pireas, Greece were analyzed in formalin fixed paraffin embedded sections. Tumors were classified according to the current World Health Organization typing of lung tumors (30) and the staging was done according to the new international staging system for lung cancer (31). No patient had received treatment of any type for their bronchial

carcinomas. Pathology of the 46 surgical specimens showed that 20 were adenocarcinomas and 26 squamous cell carcinomas.

Immunohistochemical analysis. The following methods were employed. For the detection of ras p21 the ABC method was used. The rat monoclonal antibody Y13-259 recognizing the ras p21 proteins (32) was used for the immunohistochemical analysis of the ras p21 protein. Tissue sections 5µm thick were mounted on slides and deparaffinised. Endogenous peroxidase activity was blocked by immersing the sections for 30 min in an aqueous solution of 3% H₂O₂ in the dark. The sections were washed with PBS and treated with the Y13-259 rat monoclonal antibody, goat anti-rat IgG, streptavidinperoxidase and DAB sequentially as previously described (33). Chinese hamster lung (CHL) cells, which express ras p21 at very low levels and the transformed cell line FHO5T1, which contains the mutated T24 H-ras1 oncogene inserted in a high expression vector (34) were used as controls.

Immunohistochemical analysis of the p53 protein was undertaken using the mouse anti-p53 monoclonal antibody pAb 240 (Ab-3 from Oncogene Science). pAb 240 does not bind to normal (wild type) p53 protein but recognizes a common conformational epitope on mutant p53 proteins which results from different activating mutations (35). The PAP method was used for the immunohistochemical analysis of the p53 protein. Tissue sections 5µm thick were mounted on slides and deparaffinised. Endogenous peroxidase activity was blocked by immersing the sections for 30 min in an aqueous solution of 3% H₂O₂ in the dark. The sections were washed with PBS and treated with the pAb 240 mouse anti-p53 monoclonal antibody dissolved in 5 volumes of PBS buffer and 5 volumes of bovine serum albumin 1% in ddH₂O for 1 h at 37°C in a humidified atmosphere. The slides were treated sequentially as follows: they were washed twice with PBS for 5 min each, treated with rabbit anti-mouse IgG conjugated with peroxidase 1:10 in 5 volumes PBS and 5 volumes normal human serum, incubated for 30 min at 37°C in a humidified atmosphere, washed twice with PBS for 5 min each, treated with swine anti-rabbit IgG conjugated with peroxidase 1:10 in 5 volumes PBS and 5 volumes normal human serum for 30 min at 37°C in a humidified atmosphere and washed twice with PBS for 5 min each. For localisation of the primary antibody 1 mg/ml of 3,3'- Diaminobenzidine tetra-hydrochloride solution was used. The sections were developed for 10 min at room temperature and then counterstained with Harris Hematoxylin. Control slides omitting the first antibody were used as negative controls in the immunohistochemical analysis of the p53 protein. The CM-1 antibody (36) was used with 15 specimens using the ABC technique as previously described (37). Two cell lines were used as control for p53 immunohistochemistry: the spontaneously immortalized rat 208F cells were used as negative controls for p53 expression and their transfected derivative RFV53HO6-3 cells, which carry the mutant mouse p53 gene carrying valine instead of alanine at amino acid 135, were used as positive controls. RFV53HO6-3

cells were derived after co-transfecting with the plasmid LTRp53cG-val containing the mutant p53 gene (38) and Homer 6 (34).

The immunostained sections for *ras* p21 or p53 were scored as (-/+) negative or equivocal; (+) moderate; (++) intense.

K-ras mutations.

DNA samples. DNA from 15 histological slides, fixed by formol and embedded in paraffin, was extracted by boiling in a lysis mixture.

Oligonucleotides primers and probes. The oligonucleotides were synthesized by the solid phase triester method. The primers were designed to introduce base substitution in the amplified fragments (39).

Polymerase chain reaction. In vitro enzymatic DNA amplification (PCR) was performed on an automated apparatus (DNA thermal Cycler from Perkin Elmer Cetus). We performed 35 cycles of amplification. Each cycle consisted of 3 steps: (i) denaturation of DNA at 94°C for 30 sec. (ii) annealing of the primers at 53°C for 30 sec. (iii) enzymatic extension at 72°C for 1 min.

Modified primers were designed to introduce a base substitution adjacent to the codon of interest in order to create an artificial restriction site with only one allelic form (wild type or mutated). We performed PCR with a modified primer creating a Msp I recognition site only if codon 12 was of the wild type. This approach allowed us to screen for point mutations at codon 12 of K-ras oncogene.

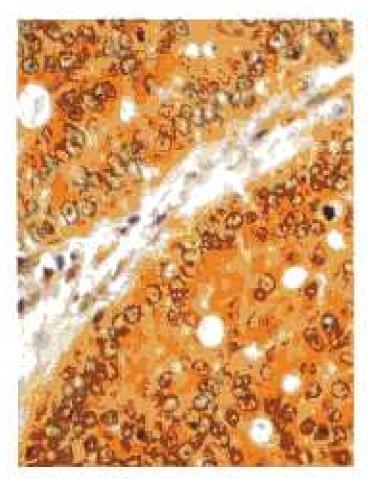
An aliquot of the PCR product was examined in a 2-3% Nu Sieve gel for the presence of the amplified fragment (99 bp). The PCR products were digested by Msp I enzyme which gave two fragments of 21 and 78 bp in all the 15 tumor DNA samples tested (all of them were wild type for codon 12 of the K-ras oncogene).

Statistical analysis. Quantitative data were analyzed by χ^2 , Fisher's exact test or McNemar's χ^2 , where appropriate. Survival curves were drawn up using the Kaplan-Meier product limit estimate (40). Differences between survival times were analyzed by the log rank method (41).

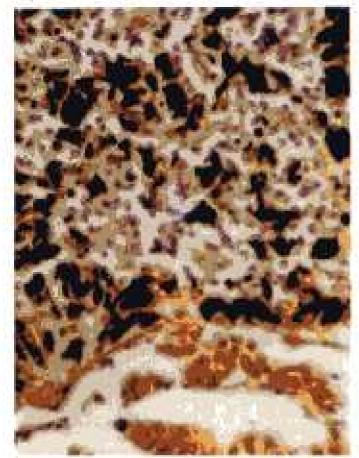
Results

Forty-six NSCLC tumors were investigated for *ras* p21 and p53 expression. The *ras* p21 monoclonal antibody Y13-259 demonstrated good cytoplasmic staining in the positive scored tumor cells, and no staining in the normal cells (Fig. 1-4). The monoclonal antibody against p53, pAb 240 gave mainly diffuse staining in a large number of the positively stained tumor cells (Fig. 5-8). Similar staining patterns were found using the CM-1 antibody in 80% of cases. This result is consistent with the finding that CM-1 recognizes both wild type and mutant p53 protein.

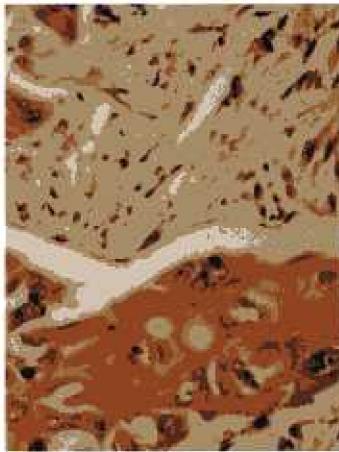
Ras p21 expression was elevated in 85% of the NSCLC tumors investigated. 90% of the adenocarcinomas and 81% of the squamous cell lung carcinomas had positive staining (Table I). Fifty-seven per cent of the NSCLC stage I; 94% of



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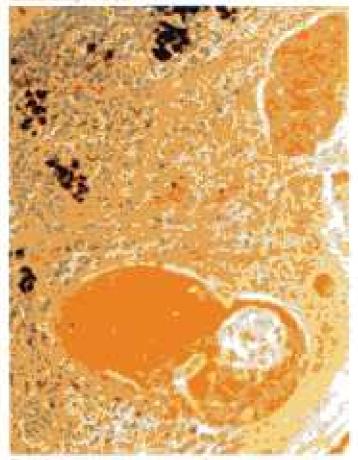


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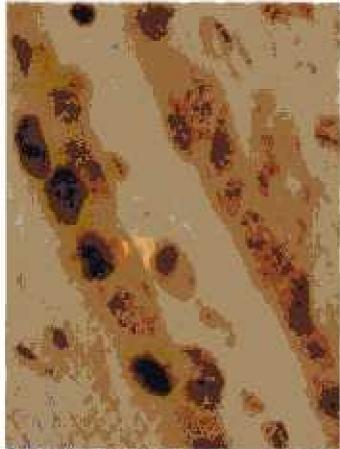


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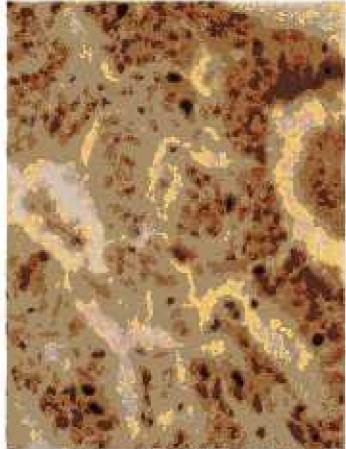
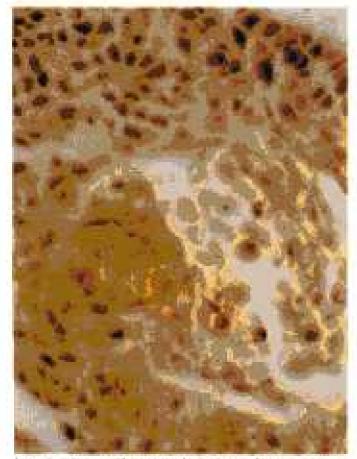


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Table I. Ras and p53 expression in non-small cell lung carcinomas.

	Total No	Intensity of st	of cases (%)	Intensity of staining/No of cases (%)			
		. /1	ras p21			p53	
Tumor type		· -/+	+	++	-/+	+	++
NSCLC ^a	46	7	17 (37)	22 (48)	25	16 (35)	5 (11)
Adenocarcinoma	20	2	9 (45)	9 (45)	11	6 (30)	3 (15)
Squamous cell carcinoma	26	5	8 (31)	13 (50)	14	10 (38)	2 (8)

^a Non Small Cell Lung Carcinoma.

Table II. Ras p21 and p53 expression in tumor and lymph node metastasis of surgically treated NSCLC patients according to stage.

	Total	Intensi	Tumor ty of staining/No	of cases (%	%)	Inten	Lymph sity of staining/		; (%)
Stage	No	ras	p21	p5	1 3	ra	s p21	ŗ	53
		-	+	-	+	-	+ '	-	+
I	14	6	8 (57)	12	2 (14)	14	0 (0)	14	0 (0)
II	18	1	17 (94)	10	8 (44)	1	17 (94)	15	3 (17)
IIIa	14	0	14 (100)	3	11 (79)	4	10 (71)	5	9 (64)
	exact tests.			1 1 1 .					
Tumor da		Τ\	•	ph node dat					
ras p<0.0	nd stage (II+II	1)		e I and Stag ><0.001	e (11+111)				
p53 p<0.0			-	p<0.001					
Stage (I+ ras p<0.0 p53 p<0.0		II	ras p	e (I+II) and >0.05 p<0.001	stage III				

stage II and 100% of stage IIIa stained *ras* p21 positive (Table II). 80% of stage I, 88% stage II and 100% of stage IIIa adenocarcinomas stained positive whereas 44% of stage I, 100% of stage II and 100% of stage IIIa squamous cell lung carcinomas stained *ras* p21 positive (Table III).

p53 expression was elevated in 46% of the NSCLC investigated. 45% of the adenocarcinomas and 46% of the squamous cell lung carcinomas stained positive with the pAb 240 monoclonal antibody (Table I). 14% of NSCLC stage I, 44% stage II and 79% stage IIIa tumors stained positive for p53 (Table II). 20% of the stage I, 50% of stage II and 57% of stage IIIa adenocarcinomas stained positive for p53 whereas 11% of stage I, 40% of stage II and 100% of stage IIIa squamous cell carcinomas stained positive for pAb 240 (Table III).

We also studied *ras* p21 and p53 expression in lymph nodes from surgically treated NSCLC patients. *Ras* p21 was over-expressed in 0% stage I; 94% stage II and 71% stage IIIa, while p53 was over-expressed in 0% stage I, 17% stage II and 64% stage IIIa in the lymph node specimens (Table II). Fifteen cases of benign lung lesions were also investigated for *ras* p21 and p53 expression; all were found to be negatively stained (Table IV).

Ras p21 and p53 staining in NSCLC were statistically analyzed with regard to staging of the tumor and lymph node specimens (Table II). Due to the small number of patients in certain categories, the staging data was analyzed by dividing them into two different groups; stage I and stage (II and IIIa), and stage (I+II) and stage IIIa. Ras p21 and p53 staining was found to be significantly different in

Table III. Ras p21 and p53 staining in tumor and lymph node metastasis of adenocarcinomas and squamous cell carcinomas of the lung according to tumor stage.

TUMOR	Total	Intens	Adenocare ity of staining		cases (%)	Total	_	mous cell car of staining/l		
Stage	rojai No	ras p21		p53		No	ras p21		p53	
Huge	7.0	-	+	-	+-		-	+	-	+
	5		.4 (80)	4	1 (20)	9	5	4 (44)	8	1 (11)
11	8	1	7 (88)	4	4 (50)	10	0	10 (100)	6	4 (40)
Ша	7	0	7 (100)	3	4 (57)	7	0	7 (100)	0	7 (100)
LYMPH	NODE MEI	ASTASIS								
ī	5	5	0 (0)	5	0 (0)	9	9	0 (0)	9	0 (0)
11	8	1	7 (88)	7	1 (13)	10	0	10 (100)	8	2 (20)
IIIa	7	2	5 (71)	5	2 (29)	7	2	5 (71)	0	7 (100)

Fisher's exact test: Stage I and stage (II and IIIa); rav (p<0.05) p53 (p<0.05)

Table IV. Immunohistochemical analysis of ras p21 and p53 protein in benign lung lesions.

		Intensity of staini	ng/No of cases	Intensity of st	aining/No of cases
		ras ţ	521	p:	53
Type of	Total				
benign lesion	No.	-	+	***	+
Echinococcus	5	5	0	5	0
Pneumonia	4	4	0	4	0
Amartoma	3	3	0	3	0
Cyst	1	1	0	1	0
Necrosis	Ī	1	0	1	0
Granulomatous tissue	1	1	0	1	0

the subgroups stage (I+II) and stage IIIa for both tumor and lymph node tissue. However, ras p21 staining was only found to be significantly different in the tumor specimens. These data indicate that there is a relationship between increasing stages of the disease with both ras and p53 expression. When the data is further subdivided into adenocarcinomas and squamous cell carcinomas (Table III), it can be seen that a similar overall relationship exists between both ras p21 and p53 staining and stage of the disease.

Ras p21 and p53 staining was also analyzed in the NSCLC according to grade of differentiation (Table V). No statistically significant correlations were found. Also when subdivided into adenocarcinomas and squamous cell carcinomas no correlations were found between stage and grade of the NSCLC specimens analyzed in this study (Table VI). No correlations were found

between ras p21 and p53 expression in this group of NSCLC patients (Table VII).

The patients' smoking history and the duration of smoking data was available for 45 or these NSCLC patients (Table VIII). All but two of these patients were either moderate or heavy smokers, the smoking duration fell into a range 10-60 years. No correlation was found between the NSCLC patients history of smoking (Table IX) or with duration (Table VIII).

Survival analysis. Stage of the disease in the NSCLC patients was found to correlate with a poor prognosis (χ^2 =29.8, df=2 p<0.001). No correlation was found for grade of differentiation with survival (χ^2 =0.65, df=2, p>0.05).

Ras p21 expression was found to be associated with survival (p<0.1) in both the tumor (χ^2 =3.6, p<0.1) (Fig. 9) and lymph node specimens (χ^2 =3.1, p<0.1) (Fig. 10). However, p53

Table V. Ras p21 and p53 expression in NSCLC according to grades of differentiation.

			Intensity of	staining,	/No of	case
	Total		ra	s p21	p:	53
Grade	No.		-	+	-	+
Low		13	3	10	. 9	4
Modeare		30	5	25	15	15
High		3	0	3	2	1

Fisher's exact test: ras p21 (p>0.05); p53 (p>0.05)

Discussion

We have examined 46 NSCLC primary tumor and nodal tissue specimens for ras p21 and p53 expression. Eighty-five per cent of these tumors expressed ras p21 with the Y13-259 antibody whereas 46% demonstrated elevated p53 mutant protein using pAb 240 monoclonal antibody. It is of particular note that elevated p53 expression correlated with survival in both the tumor (p<0.05) and also in lymph node metastasis specimens (p<0.005). This is the first publication that we know of which indicates a correlation between mutant p53 expression and a poor prognosis in NSCLC patients. The over-expression of ras p21 in these tumors was also found to be associated with a poor survival but was not statistically significant. However, Miyamoto et al (42) have demonstrated a correlation

Table VI. Ras p21 and p53 staining in adenocarcinomas and squamous cell carcinomas of the lung according to grades of differentiation.

		A	denocar	cinom	as		Sc	luamous (cell carci	nomas	
		Intensity	of stain	ing/N	o of cases		Inten	sity of sta	ining/No	o of cases	
Grade	Total	ras	s p21	p:	53	Total	<i>ras</i> p21		p:	p53	
	No	-	+	-	+	No	-	+	-	+	
Low	6	0	6	4	2	7	3	4	5	2	
Moderate	13	3	10	7	6	17	2	15	8	9	
High	1	0	1	1	0	2	0	2	1	1	

Table VII. Interrelationship between the expression of ras p21 and p53 in 46 NSCLC patients.

Intensity of staining/ No of cases	Intensity of staining/No of cases							
		ras	p21					
		-	+					
p53	-	5	20					
•	+	2	19					

McNemar's χ^2 test; ras p21 and p53 staining results of the tumor data p<0.001

over-expression was found to correlate with a poor prognosis in the tumor specimens (χ^2 =3.9, p<0.05) (Fig. 11) and was found to be highly significant in lymph node tissues (χ^2 =10.6, p<0.005) (Fig. 12). No correlation was found between survival and smoking in these NSCLC patients.

Mutations in the K-ras gene. Mutations in K-ras, codon 12 has been assessed in a further group of 15 NSCLC patients, 6 adenocarcinomas, and 9 squamous cell carcinomas. None of these patients demonstrated a mutation in K-ras codon 12.

between ras p21 and survival using the ras rp-35 monoclonal antibody in a large survey of 112 NSCLC patients.

A number of clinico-pathological parameters including stage, grade of differentiation as well as the patient's smoking history were analyzed, in relation to the expression of *ras* p21 and p53 proteins.

In this study, stage of the disease was shown to correlate with a poor prognosis, as p53 and also *ras* p21, to some degree, correlated with survival, it is not surprising that the over-expression of these two genes correlated with the stage of the disease (Table II). This correlation was found in both the tumor and lymph node tissue data. On subdividing the NSCLC into squamous cell carcinomas and adenocarcinomas this association was again found (Table III). No association was found for either *ras* p21 or p53 expression with the grade of differentiation of the NSCLC (Table V).

In this investigation we have demonstrated *ras* p21 over-expression in 81% of the squamous cell carcinomas compared to 90% of the adenocarcinomas. Spandidos *et al* (17) have previously reported significant differences between these two histological groups of NSCLC tumors. Similar findings were obtained from Rodenhuis *et al* (10) and Shiraishi *et al* (43) but not Kurzrock *et al* (44) and Koutselini *et al* (18) who found increased levels of *ras* p21

Table VIII. Ras p21, p53 clinicopathological parameters, smoking history and follow up on the 46 NSCLC patients analyzed in this study.

Pat.	Stage	Type	Grading	Sex		ng history	Follow up	Fate
No.	C	• •			Type	Duration	(months)	
1	I	AD	moderate	m	Н	40	52	A
2	I	SQ	low	m	Н	40	47	Α
3	I	AD	moderate	m	Н	35	52	Α
4	I	SQ	moderate	m	M	25	39	Α
5	I	SQ	low	m	M	30	38	Α
6	I	SQ	moderate	m	Н	30	26	Α
7	I	AD	low	m	Н	20	13	D
8	I	SQ	low	m	Н	30	26	Α
9	I	SQ	moderate	m	M	25	30	Α
0	I	SQ	moderate	m	Н	40	50	A
1	I	AD	moderate	m	M	50	23	D
2	I	AD	moderate	m	M	43	28	Α
3	I	SQ	low	m	Н	20	48	Α
4	I	SQ	moderate	m	Н	50	14	D
5	II	AD	moderate	m	Н	50	26	D
6	II	SQ	moderate	m	Н	45	10	D
.7	II	SQ	moderate	m	M	45	28	Α
8	II	AD	moderate	m	Н	30	36	Α
9	II	SQ	moderate	m	Н	30	42	Α
0	II	SQ	high	m	Н	30	36	Α
21	II	AD	moderate	m	ND	ND	23	D
2	II	SQ	high	m	Н	50	35	Α
23	II	AD	low	m	Н	50	30	D
24	II	SQ	low	m	M	50	25	D
25	П	SQ	moderate	m	Н	38	23	D
26	II	SQ	moderate	m	N	ND	36	Α
27	II	SQ	low	m	Н	50	48	Α
28	II	AD	low	m	Н	50	60	A
9	II	AD	high	m	M	50	38	D
80	II	SQ	moderate	m	Н	30	30	A
31	II	AD	moderate	m	H	20	60	A
32	II	AD	moderate	m	M	60	39	D
33	IIIa	SQ	moderate	m	M	40	20	D
34	IIIa	SQ	moderate	m	Н	40	6	D
35	IIIa	SQ	low	m	M	25	18	D
36	IIIa	AD	moderate	m	M	40	5	D
37	IIIa	AD	low	f	N	ND	24	D
38	IIIa	AD	moderate	m	M	10	26	A
9	IIIa	AD	moderate	m	H	20	4	D
10	IIIa	SQ	moderate	m	Н	40	18	D
1	IIIa	AD	low	m	M	40	12	D
12	IIIa	AD	moderate	m	Н	21	6	D
13	IIIa	AD	low	m	H	30	18	D
14	IIIa	SQ	moderate	m	Н	25	11	D
15	IIIa	SQ	moderate	m	M	30	6	D
16	IIIa	SQ	moderate	m	M	40	17	D

Stage: New International Staging System for lung cancer (31). Type of carcinoma: AD = adenocarcinoma; SQ = Squamous cell lung carcinoma. Sex: m = male; f = female. Smoking type: non smoker (N); moderate smoker (M) (less than 20 per day); heavy smoker (H) (greater than 20 per day). Duration: duration of smoking habit in years. Fate: A = Alive; D = Dead; ND = No data.

oncoprotein in tumors with a squamous carcinoma histology using the same antibody.

Ras p21 expression was found in all stages of the disease but was particularly increased in stages II and IIIa in both the tumor and nodal tissue specimens. Rodenhuis et al (6) demonstrated that the adenocarcinomas with K-ras mutations tended to be in smaller tumors that had less often spread to the regional lymph nodes (6,10,12). The results of this

investigation would indicate that *ras* over-expression was a late event in the progression of NSCLC patients.

In the present investigation p53 staining was found in 46% of NSCLC using the pAb 240 monoclonal antibody against the mutant p53 protein. This finding agrees with that of Hiyoshi *et al* (45) who used the p53 pAb 1801 antibody. They demonstrated p53 over-expression in 57% of the squamous cell carcinomas and in 43% of the adenocarcinomas. It is also

Table IX.	Relationship between	NSCLC patients	smoking history	and ras p21 ar	nd p53 expression.

		ras p21 Intensity of st				p53	3	
	Tumor		N	ode		Tumor Node		
	-	+	-	+	-	+	-	+
Non smoker	0	2	1	1	0	2	1	1
Mod. smoker	3	13	8	8	8	8	12	4
Heavy smoker	4	23	10	17	17	10	20	4

of note that Hiyoshi *et al* (45) demonstrated that p53 staining correlated with regional node metastasis, distant metastasis and pathological stage in adenocarcinomas. Furthermore, in this present investigation p53 over-expression was found to significantly correlate with the stage of the disease, particularly in the lymph node specimens (p<0.005).

The role of smoking in lung cancer has been extensively reviewed (46). However, recent evidence also indicate a genetic link between the K-ras, p53 and a history of

smoking. Rodenhuis *et al* (6) demonstrated that K-ras mutations were found in about one third of the adenocarcinomas of the lung investigated and that these mutations correlated with the patients smoking history. Furthermore, no other clinical correlations were noted with the K-ras mutations except the patient's smoking history. None of the non-smokers in their study had a mutation whereas 13 of 32 smokers did have a K-ras mutation. Similar results were found by Slebos *et al* (47). Kobayashi *et al* (7) have found a

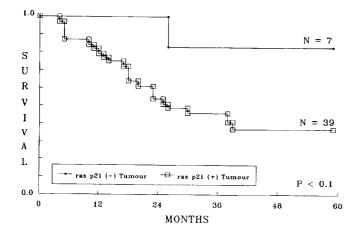


Figure 9. Survival curve of NSCLC tumor tissue for *ras* p21 expression, drawn up by Kaplan Meier (40) and log rank calculated by Peto (41).

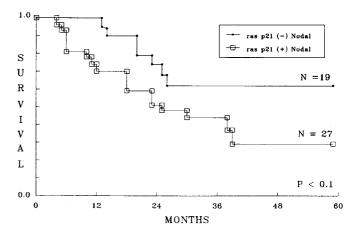


Figure 10. Survival curve of NSCLC nodal metastasis of surgically treated NSCLC patients for *ras* p21 expression, drawn up by Kaplan Meier (40) and log rank calculated by Peto (41).

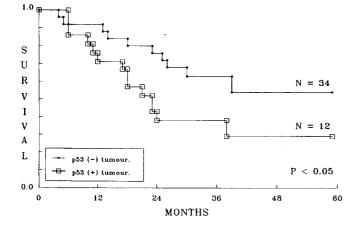


Figure 11. Survival curve of NSCLC tumor tissue for p53 expression, drawn up by Kaplan Meier (40) and log rank calculated by Peto (41).

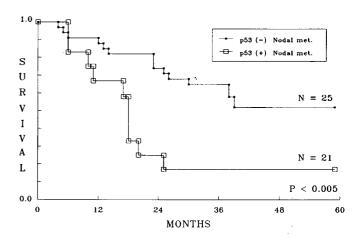


Figure 12. Survival curve of NSCLC nodal metastasis of surgically treated NSCLC patients for p53 expression, drawn up by Kaplan Meier (40) and log rank calculated by Peto (41).

higher incidence of K-ras mutations in NSCLC, than in SCLC and a correlation was found between these mutations and smoking habits in the non-goblet all types of these tumors. We also investigated the smoking history of the 45 NSCLC patients in this study, however, as only two patients were non-smokers it is not surprising no correlations were forthcoming.

The evidence associating p53 mutations and a history of smoking has become more compelling. Chiba et al (8) demonstrated that 56% of p53 mutations in the lung tumors were G to T transversions unlike many other tumors. It is believed that the type of mutation is usually associated with a specific mutagen, for example benzo (a) pyrene may cause G to T transversions in certain circumstances (48) whereas different mutagens can cause G to A transversions. This was the first molecular evidence which suggested that lung cancer is caused by a specific mutagen and may indicate that a particular carcinogen in the smoke causes lung cancer. Although Chiba et al (8) did not demonstrate a correlation between p53 mutations and a history of smoking, this data has been recently provided by a Japanese group (9) which clearly found a correlation between a lifetime cigarette consumption and p53 mutations in lung cancer patients. It is also of particular interest that Field et al (49,50) have found a correlation of the head and neck cancer and a history of heavy smoking and drinking.

When these results are considered together, they suggest that the p53 gene may play a role in the early development of lung cancer. Since life-time cigarette consumption correlated with p53 mutations (9) and nodal metastasis, stage and poor prognosis also correlates with p53 overexpression (our data), it may be proposed that the aberrant expression of this gene has also a role in the late events in lung neoplasias.

References

- Minna J, Pass H, Glatstein E and Ihde D: In: Cancer: Principles and Practice of Oncology. 3rd Devita J, VT Hellman S and Rosenberg S: (eds) JB Lippincott Co Philadelphia. pp591- 705, 1989.
- Minna JD, Ihde DC, Glatstein EJ: Lung cancer: scalpels, beams, drugs and probes. (editorial) N Engl J Med 315: 1411-1414, 1986.
- 3. Minna JD, Schütte J, Viallet J, Thomas F, Kaye FJ, Takahashi T, Nau M, Whang-Peng J, Birrer M, Gazdar AF: Transcription factor and recessive oncogenes in the pathogenesis of human lung cancer. Int J Cancer Suppl 4: 32-34, 1989.
- 4. Field JK and Spandidos DA: The role of *ras* and *myc* oncogenes in human solid tumors and their relevance in diagnosis and prognosis. Anticance: Res 10: 1-22, 1990.
- prognosis. Anticancei Res 10: 1-22, 1990.

 5. Harris CC: Chemical and physical carcinogenesis: advances and perspectives for the 1990s. Cancer Res (Suppl) 51: 5023S-5044S, 1991.
- Rodenhuis S, Slebos R, Boot A. Evers S, Mooi W, Wagenaar S, Van Bodegom P and Bos J: Incidence and possible clinical significance of K-ras oncogene activation in adenocarcinoma of the human lung. Cancer Res 48: 5738-5741, 1988.

7. Kobayashi T, Tsuda H, Noguchi M, Hirohashi S, Shimosato Y, Goya T, Hayata Y: Association of point mutation in c-Ki-ras oncogene in lung adenocarcinoma with particular reference to cytologic subtypes. Cancer 66 (2): 289-294, 1990.

8. Chiba I, Takahashi T, Nau M, D'Amico D, Curiel D, Mitsudomi T, Buchhagen D, Carbone D, Piantadosi S, Koga H, Reissmann P, Slamon D, Holmes EC and Minna J: Mutations in the p53 gene are frequent in primary resected non-small cell lung cancer. Oncogene 5: 1603-1610, 1990.

 Suzuki H, Takahashi T, Suyama M, Arioshi Y, Takahashi T and Ueda R: p53 mutations in non-small cancer cell cancers in Japan: Association between mutations and smoking. Cancer Res 52, 24, 724, 1002

52: 734-736, 1992.

- Rodenhuis S, Van de Wetering M, Mooi W, Evers S, Van Zandwijk N and Bos J: Mutational activation of the K-ras oncogene: a possible pathogenetic factor in adenocarcinoma of the lung. N Engl J Med 317: 929-935, 1987.
- 11. Suzuki Y, Orita M, Shiraishi M, Hayashi K and Sekiya T: Detection of *ras* gene mutations in human lung cancers by single-strand conformation polymorphism analysis of polymerase chain reaction products. Oncogene 5: 1037-1043, 1990.
- 12. Slebos RJC, Kibbelaar RE, Dalesio O, Kooistra A, Stam J, Meijer CJLM, Wagenaar SS, Vanderschueren RGJRA, Van Zandwijk N, Mooi WJ, Bos JL and Rodenhuis S: K-ras oncogene activation as a prognostic marker in adenocarcinomas of the lung. N Engl J Med 323: 561-565, 1990.
- 13. Mitsudomi T, Steinberg SM, Oie HK, Mulshine JL, Phelps R, Viallet J, Pass H, Minna JD and Gazdar AF: Ras gene mutations in non-small cell lung cancers are associated with shortened survival irrespective of treatment intent. Cancer Res 51: 4999-5002, 1991.
- Venenzuela DM and Groffen J: Four human carcinoma cell lines with novel mutations in position 12 of c-K-ras oncogene. Nucleic Acid Res 14: 843-852, 1986.
- Milici A, Blick M, Murphy E and Gutterman JU c-K-ras codon 12 GGT-CGT point mutation: an infrequent event in human lung cancer. Biochem Biophys Res Commun 140: 669-705, 1986.
- 16. Nakano H, Yamamoto F, Neville C, Evans D, Mizuno T and Perucho M: Isolation of transforming sequences of two human lung carcinomas: structural and functional analysis of the activated c-K-ras and oncogenes. Proc Natl Acad Sci USA 81: 71-75, 1984.
- 17. Spandidos DA, Zakinthinos S, Petraki C, Sotsiou F, Yiagnisis M, Dimopoulos AM, Roussos C and Field JK: Expression of *ras* p21 and *myc* p62 oncoproteins in small cell and non-small cell carcinoma of the lung. Anticancer Res 10: 1105-1114, 1990.
- 18. Koutselini H, Kappatou G, Yiagnisis M, Field JK and Spandidos DA: Immunohistochemical study of RAS oncoprotein in cytologic specimens of primary lung tumors. Anticancer Res 10: 597-604, 1990.
- 19. Hollstein M, Sidransky D, Vogelstein B, Harris CC: p53 mutations in human cancers. Science 253: 49-53, 1991.
- 20. Bartek J, Bartkova S, Vojtesek B *et al*: Aberrant expression of the p53 oncoprotein is common feature of a wide spectrum of human malignancies. Oncogene 6: 1699-1703, 1991.
- 21. Farmer G, Burgonetti J, Zhu H, Friedman P, Prywes R and Prives C: Wild-type p53 activates transcription *in vitro*. Nature 358: 83-86, 1992.
- Oliner JD, Kinzler KW, Meltzer PS, George DL and Vogelstein B: Amplification of a gene encoding a p53associated protein in human sarcoma. Nature 358: 80-83, 1992.
- 23. Lane DP: p53, guardian of the genome. Nature 358: 15-16, 1992.
- 24. Baker SJ, Fearon ER, Nigro JM, Hamilton SR, Preisinger AC, Jessup JM, Van Tuinen P, Ledbetter DH, Barker DF, Nakamura Y, White R and Vogelstein B: Chromosome 17 deletions and p53 gene mutations in colorectal carcinomas. Science 244: 217-221, 1989.
- 25. Takahashi T, Takahashi T, Suzuki H, Hida T, Sekido Y, Ariyoshi Y and Ueda R: The p53 gene is very frequently mutated in small cell lung cancer with a distinct nucleotide substitution pattern. Oncogene 6: 1775-1778, 1991.
- Miller CW, Simon K, Aslo A, Kok K, Yokota J, Buys CM, Terada M and Koeffler HP: p53 mutations in human lung tumors. Cancer Res 52: 1695-1698, 1992.
- 27. Iggo R, Gatter K, Bartek J, Lane D and Harris AL: Increased expression of mutant forms of p53 oncogene in primary lung cancer. Lancet 335: 675-679, 1990.
- 28. Nigro JM, Baker SJ, Preisinger AC, Jessup JM, Hostetter R, Cleary K, Bigner SH, Davidson N, Baylin S, Devilee P, Glover T, Collins FS, Weston A, Modali R, Harris CC and Vogelstein B: Mutations in the p53 gene occur in diverse human tumor types. Nature 342: 705-708, 1989.
- 29. Bodner SM, Minna JD, Jensen SM, D'Amico D, Carbone D, Mitsudomi T, Fedorko J, Buchhagen L, Nau MN, Gazdar AF and Linnoila I: Expression of mutant p53 proteins in lung cancer correlates with the class of p53 gene mutation. Oncogene 743-749, 1992.
- 30. World Health Organization: Histological typing of lung tumors. 2nd ed. Am J Clin Path 77:123-136, 1982.
- 31. Mountain CF: A new international staging system for lung cancer. Chest 89: 225S-233S, 1986.

- 32. Furth ME, Davis IJ, Fleurdely B and Scolnick EM: Monoclonal antibodies to the p21 products of the transforming gene of Harvey murine virus and of the cellular gene family. J Virol 43: 294-304, 1982.
- 33. Papadimitriou K, Yiagnisis M, Tolis G and Spandidos DA: Immunohistochemical analysis of the *ras* oncogene product in human thyroid neoplasms. Anticancer Res 8: 1223-1228, 1988.
- 34. Spandidos DA and Wilkie NM: Malignant transformation of early passage rodent cells by a single mutated human oncogene. Nature 310: 469-475, 1984.
- 35. Gannon JN, Greaves R, Iggo R and Lane DP: Activating mutations in p53 produce a common conformational effect. A monoclonal antibody specific for the mutant form. EMBO J 9: 1595-1602, 1990.
- Midley CA, Fisher CJ, Bartek J, Vojtesek B, Lane DP, Barnes DM: Analysis of p53 expression in human tumors: an antibody raised against human p53 expressed in E coli. J Cell Science 101: 183-189, 1992.
- 37. Field JK, Malliri A, Jones AJ and Spandidos DA: Mutations in the p53 gene at codon 249 are rare in squamous cell carcinoma of the head and neck. Int J Oncol 1: 253-256, 1992.
- 38. Finlay CA, Hinds PW and Levine AJ: The p53 proto-oncogene can act as a suppressor of transformation. Cell 57: 1083-1093, 1989
- 39. Skalkeas Gr, Spandidos DA, Kostakis A, Balafouta-Tseleni S, Choremi E, Iliopoulos D, and Haliassos A: K-ras oncogene mutations in neoplasias of kidney transplanted patients: preliminary results with a new technique. Anticancer Res 11: 2091-2094, 1991.
- Kaplan EL and Meier P: Non-parametric estimation from incomplete observations. J Amer Stat Assoc 53: 457-481, 1958.
- 41. Peto R, Pike MC, Armitage PE, Breslow NE, Cox DR, Howard SV, Mantel N, McPherson K, Peto J and Smith PG: Design and analysis of randomized clinical trials requiring prolonged observations of each patient. Br J Cancer 34: 585-612, 1976.

- 42. Miyamoto H, Harada M, Isobe H, Akita HD, Haneda H, Yamaguchi E, Kuzumaki N, Kawakami Y: Prognostic value of nuclear DNA content and expression on the *ras* oncogene product in lung cancer. Cancer Res 51: 6346-6350, 1991.
- 43. Shiraishi M, Noguchi M, Shimosato Y and Sekiya T: Amplification of proto-oncogenes in surgical specimens of human lung carcinomas. Cancer Res 49: 6474- 6479, 1989.
- 44. Kurzrock R, Gallick GE and Gutterman JU: Differential expression of p21 *ras* gene products among histological subtypes of fresh primary human lung tumors. Cancer Res 46: 1530-1534, 1986.
- 45. Hiyoshi H, Matsuno Y, Kato H, Shimosato Y, Hirohashi S: Clinicopathological significance of nuclear accumulation of tumor suppressor gene p53 product in primary lung cancer. Jpn J Cancer Res 83 (1): 101-106, 1992.
- Lilienfeld AM and Lilienfeld DE: Foundation of epidemiology. New York, Oxford University Press, 1980.
- Slebos RJ, Hruban RH, Dalesio O, Mooi WJ, Offenhaus GJ, Rodenhuis S: Relationship between K-ras oncogene activation and smokin in adenocarcinomas of the human lung. J Natl Cancer Inst 83: 1024-1027, 1991
- Cancer Inst 83: 1024-1027, 1991.

 48. Mazur M and Glikman B: Sequence specificity of mutations induced by benza[alpha] pyrene-7,8 ohol-9,10 epoxide at endogenous part gene in CHO cells. Som cell. Mol Gen 14: 393-400, 1988.
- 49. Field JK, Spandidos DA, Malliri A, Gosney JR, Yiagnisis M and Stell PM: Elevated p53 expression correlates with a history of heavy smoking in squamous cell carcinoma of the head and neck. Br J Cancer 64: 573-577, 1991.
 50. Field JK, Spandidos DA and Stell PM: Overexpression of the
- 50. Field JK, Spandidos DA and Stell PM: Overexpression of the p53 gene in head and neck cancer a genetic link with heavy smoking and drinking. Lancet 339: 502-503, 1992.