The role of oncogenic kinases in human cancer (Review)

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Received March 16, 2000; Accepted April 14, 2000

Abstract. Tumorigenesis in humans is a multistep process, which reflects genetic alterations that lead to cell transformation and malignancy. Cellular genes that are altered are normally involved in maintaining cell homeostasis by participating in signaling pathways tightly regulated to maintain the functional integrity of the cell. When these genes are altered they escape from the regulatory control and transmit signals that lead to the progressive conversion of normal cells into cancer cells. Oncogenic signals involve activation of kinases, which can be either a primary event when they are directly mutated in a tumor cell or a secondary event as recipients and mediators of oncogenic signals. Transmembrane (e.g. EGFR, PDGFR) or cytoplasmic (Src, Abl) tyrosine kinases are found mutated in a variety of human tumors. Cytoplasmic serine threonine kinases (Raf, Akt, Tpl-2) are also mutated or activated in several types of human malignancies. Kinases transduce signals that lead to cell proliferation or inhibition of programmed cell death by activating transcription factors (e.g. AP1, NFkB, Myc), inhibiting pro-apoptotic molecules (e.g. Bad, Bax), or they participate in deregulating the cell cycle control. Thus, kinases play a central role in oncogenesis rendering them putative targets for anti-cancer drug design.

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Key words: human cancer, tyrosine kinases, serine threonine kinases, apoptosis, Abl, PI3 kinase, Akt, Raf, Tpl-2, Ets, NFκB, myc, CDK, Rb

1. Introduction

Cancer arises when one or more cells lose their ability to control cell division and they begin to proliferate in an uncontrolled fashion. The origin of cancer lies in the genetic material of the cell and is a result of an accumulation of mutations that promote clonal selection of cells with an aggressive phenotype. This phenotype is underlined by the faster proliferation rate and alterations in the cell morphology. To find effective therapeutic interventions for cancer we need to understand the events that take place during cell transformation. Since cancer originates in the genetic material of the affected cell, the primary step is the identification of the genes that are altered in the tumor cell. These genes are defined as oncogenes, genes that are usually either overexpressed or mutated so that they cannot be regulated as they used to, and oncosuppressor genes, genes that normally function as brakes in the cell cycle or repair damaged DNA and when their function is lost the cell loses control of its division rate or acquires mutations that lead to faster proliferation (1,2). The second step is the understanding of the role of the proteins encoded by these genes in the cellular environment. In other words, we need to understand the function of these proteins in the normal cell and in the tumor cell. By elucidating the mechanism through which these proteins induce the tumor we can interfere with therapeutic agents that will be able either to specifically inhibit the function of the genes involved eliminating the cells, or perturb their proliferation and lead them to extinction (3).

All oncogenic proteins participate in cellular functions that involve transduction of signals from the extracellular environment, through the membrane, into the cytoplasm towards the nucleus, where transcription is initiated to generate proteins that will eventually contribute to the oncogenic phenotype. Study of the signal transduction in cancer can be therefore divided into the following areas: a) signaling from growth factors and cytokines via transmembrane receptors, b) role of the cytoplasmic signaling molecules in cancer c) regulation of transcription factors in cancer. These signaling events have effects a) on the regulation of the cell cycle and b) the regulation of apoptosis.

Cellular signaling involves phosphorylation events that occur through interactions of kinases that are localized on the cell membrane, in the cytosol or in the nucleus. Kinases that are often deregulated in human cancers are able to initiate or alter signals that eventually lead to cell proliferation and transformation. Such kinases transfer phosphates in tyrosine residues or serine and threonine residues of other kinases or

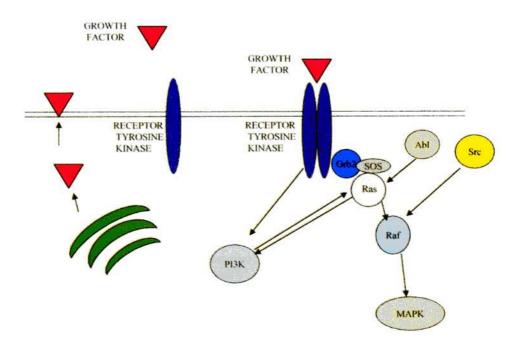


Figure 1. Signaling via transmembrane receptors. Receptor tyrosine kinases are bound to their ligands, dimerize and transmit signals that lead to proliferation and differentiation.

other proteins that participate in various signaling pathways in the cell.

2. Signaling via transmembrane receptors

Transmembrane receptors often contain domains that exhibit catalytic properties. The tyrosine kinase receptors are a major family of transmembrane receptors. Tyrosine kinases act to transfer phosphate from ATP to tyrosine residues on specific cellular proteins. Phosphorylation of these proteins alters their catalytic properties or their association with other molecules and therefore initiates biochemical signals that lead to gene transcription or morphological changes. Tyrosine kinases can be either transmembrane or cytoplasmic. The first oncogene identified was the tyrosine kinase v-src which was encoded by the Rous sarcoma virus in chicken. The cellular homologue c-src is a non-receptor kinase tightly regulated in contrast with its oncogenic viral counterpart (4,5).

Tyrosine kinase receptors can contribute to the oncogenic phenotype by different mechanisms:

a) Secretion of particular growth factors may be deregulated and as a result the receptor will be triggered at a higher than the normal level. Often tumors are found to secrete growth factors such as epidermal growth factor (EGF), colony stimulating growth factor 1 (CSF1), insulin growth factor I (IGF-I) and platelet-derived growth factor (PDGF) (6). These factors bind to their receptors and initiate growth and proliferative signals. This mechanism establishes an autocrine loop that leads to tumor growth (Fig. 1).

b) Tyrosine kinase receptors often dimerize or oligomerize following ligand binding. The dimerization and the conformational changes that are induced by ligand binding bring the cytoplasmic tails in such proximity as to trigger autophosphorylation. Autophosphorylation in most cases activates a cascade of phosphorylation events that include

phosphorylation of intracellular signaling molecules and recruitment of SH2 (*src* homology 2) domain-containing proteins that bind to specific tyrosine phosphorylated residues (7,8). In various tumors tyrosine kinase receptors can be constitutively activated by mutations that render them active independent of ligand binding. Such mutations were found on NEU/*c-erb*B-2 (9,10). Mutation of the transmembrane domain was also found in other viral oncogenes such as v-ROS, which obtains a very broad substrate specificity (11).

c) Alternatively, tyrosine kinases can become oncogenic by mutations that make them active independent of ligand binding or dimerization. Non-receptor tyrosine kinases are also activated by mutations that affect their negative regulation such as the mutation on tyrosine 527 of Src that leads to deregulation of its activation (12).

d) Several tyrosine kinases are activated in tumors via mutations. A major example is the BCR-ABL that is a mutant protein caused by the reciprocal translocation between chromosomes 9 and 22, the Philadelphia chromosome, that juxtaposes sequences of the breakpoint cluster region BCR on chromosome 22 with the c-ABL kinase on chromosome 9 (13,14). This translocation is present on 95% of chronic myelogenous leukemias, which account for 20% of the adult leukemias. The BCR-ABL fusion gene in CMLs produces a protein in which the first exon of c-ABL has been replaced by BCR sequences encoding 927 or 902 amino acids (15,16). In other cases 185 kDa BCR portion is fused with exons 2-11 of the c-ABL protein (17). The BCR-ABL chimeric protein exhibits tyrosine kinase activity several fold higher than that of the c-ABL. This kinase can transform fibroblasts and is considered highly oncogenic (18,19). The pathways through which this protein causes transformation are not clearly defined. It is known that it binds and activates GRB-2 (20) which in turn activates the Ras pathway, a key pathway for triggering MAPK activation and cell proliferation. Other

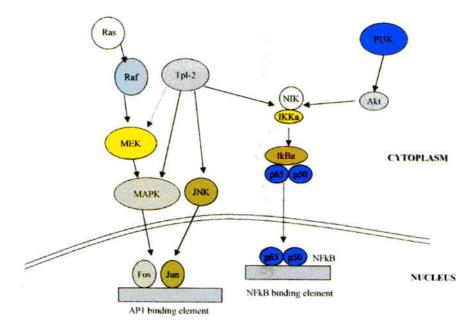


Figure 2. Signaling via cytoplasmic proteins. Activation of transcription factors. Oncogenic serine threonine kinases transduce signals to activate the MAPK, JNK and NFκB pathways which in turn promote gene expression.

known fusion proteins are TEL-ABL, present in acute lymphoblastic leukemia (ALL), in acute myeloid leukemia (AML) and in chronic myeloblastic leukemia (CML) with a reciprocal t(9;12) translocation which links the Ets-like transcription factor TEL with the ABL tyrosine kinase (21-23). TEL has also been found fused to the PDGF receptor (TEL-PDGFR) in chronic myelomonocytic leukemias (CMML) through an acquired translocation in hematopoietic cells, t(5;12)(q33;p13) (24-26).

Other receptors involved in the signaling in tumor cells are the cytokine and growth factor receptors that transduce signals from cytokines and growth factors often expressed by tumor cells, such as the TGFB in breast tumors (27). Antigen receptors also play a significant role in tumor formation either by giving the tumor cell the ability to escape the immune system surveillance or by rendering hematopoietic cells sensitive to proliferation signals.

3. Signaling via cytoplasmic proteins

The signal that is initiated at the transmembrane receptors is being transduced through cytoplasmic proteins via the cytoplasm into the nucleus. The cytoplasmic signaling molecules can be protein kinases, phosphatases or other proteins such as adaptor molecules. Most transmembrane proteins are associated with intracellular tyrosine or serine threonine kinases, which, in turn activate signaling cascades towards the nucleus.

Activation of the MAPKinase and the PI3Kinase cascades are critical events during cell activation and proliferation. Several oncogenes are known to act on these pathways and several molecules that participate on these cascades when deregulated become oncogenic. *Ras*, a well-studied family of oncogenes, structurally altered in about 25% of all human tumors, functions on activating the MAPK cascade (28-30). Raf1, a serine threonine kinase that is activated by Ras, is also found to be activated in myeloid leukemias (31,32).

Cytoplasmic oncogenes can be serine threonine kinases. In this family of oncogenes the most important ones are the Akt family (Akt1, Akt2, Akt3). Akt2 was found to be activated in pancreatic adenocarcinomas, small cell lung cancer, and ovarian cancers (33-35). Akt3 was also found activated in estrogen receptor deficient breast cancers and androgen independent prostate cancers (36). The Tpl-2/Cot oncogene is activated in breast (37), thyroid and colon tumors (38).

The Tpl-2 oncogene activates the MAPKinase (mitogen activated protein kinase) and the SAPKinase (stress activated protein kinase) pathways (39,40). Activation of these two pathways leads to the activation of transcription factors such as AP1 and NFAT (41,42). Tpl-2 also activates the transcription factor NF κ B, by activating the kinase that phosphorylates and induces degradation of the NF κ B inhibitor IkB α (43-45). Activation of these factors induces transcription of several genes that contribute to the tumor phenotype (Fig. 2).

The Akt proto-oncogene (46) is activated by PDGF receptor via activation of the PI3Kinase, a kinase that phosphorylates lipids (47,48). The lipids bind to the PH domain of Akt and induce its serine threonine kinase activity (49). Activation of Akt inhibits apoptosis by inhibiting BAD, a pro-apoptotic, Bcl-2-binding protein (50-52). Akt is also involved in inducing cell cycle progression possibly by activating transcription factors such as NFkB (53,54). Akt kinase is known to induce phosphorylation of IkB α via NIKinase and IKK α (53). It is also a transducer of growth factor signals such as PDGF, G-CSF, IL-2, hepatocyte growth factor, IGF and other mitogenic signals. Most of these signals lead to phosphorylation of Akt which results in signals that lead to inhibition of apoptosis (48,55,56). The pleiotropic effect of Akt and other oncogenic molecules is often regulated by other oncogenic molecules. Thus, when a combination of such oncogenes is activated a particular phenotype is favored. For example, in breast tumor cells Akt phosphorylates Raf at a highly conserved serine

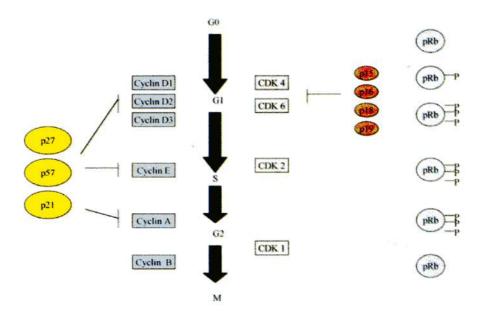


Figure 3. Cell cycle control. Progression through the cell cycle involves different regulatory mechanisms that require phosphorylation of cyclins by the CDKs and subsequent change in the phosphorylation status of the Rb protein.

residue in its regulatory domain *in vivo*. This phosphorylation of Raf by Akt inhibited activation of the Raf-MEK-ERK signaling pathway and shifted the cellular response from cell cycle arrest to proliferation (57). Such interactions can occur and determine the levels of crosstalk and fine regulation of different signaling pathways, the MAPK and the PI-3Kinase.

The examples of Raf, Akt and Tpl-2 indicate that an oncogenic serine threonine kinase can contribute to tumor formation through pleiotropic effects. On the one hand they can induce transcription of genes that are normally not expressed in these cells and on the other hand they can directly interfere with cell cycle machinery and promote progression through the cell cycle. Alternatively, they can inhibit programmed cell death and, therefore, allow the survival of a cell that carries other defects and would otherwise enter apoptosis.

4. Signaling to the nucleus

Cytoplasmic oncogenes often lead to the activation of transcription factors. The transcription factors themselves, though, can be activated by several mechanisms during tumorigenesis and contribute to tumor formation. Such signals are often regulated via phosphorylation of transcription factors either in the cytoplasm or in the nucleus. In the case of NF κ B a sequence of phosphorylation events leads to degradation of its inhibitory molecule, IkB α and its subsequent translocation into the nucleus. In other cases, such as NFAT, dephosphorylation by calcineurin leads to its nuclear translocation and a nuclear kinase, GSK3 phosphorylates NFAT which translocates into the cytoplasm (58).

In some cases a transcription factor is mutated and activated independent of extracellular or cytoplasmic signals. Expression of the transcription factors Ets-1 and Ets-2 is induced during cell proliferation but it has also been directly linked to a complex chromosomal translocation, t(6;18;21), in acute non

lymphoblastic leukemias. Ets-2 is overexpressed during hepatic regeneration and in hepatocellular carcinomas (59). In other cases activation of the signaling pathways previously mentioned lead to cell differentiation and proliferation. These events require different genes to be expressed. NFκB is a major transcription factor found to be activated in breast tumors, pancreatic adenocarcinomas, lung cancers and acute T cell leukemias (60-62). Another transcription factor involved in various human tumors is c-myc. When over-expressed in human tumors it dimerizes with Max, a complex that elicits growth signals, while the Mad-Max complex promotes differentiation signals (63-65). Overexpression of c-myc has been shown to be involved in human tumors including colon, stomach, cervix, breast and haematological neoplasms (66-69).

5. Cell cycle progression

A tumor cell is characterized by short and uncontrolled proliferation. All oncogenic events lead to deregulation of the cell machinery that controls the cell cycle. In other cases the cell cycle components have been affected and the cell loses the ability to control its proliferation. Whether the effect is direct, involving mutation of genes that regulate the cell cycle, or indirect, the result is a shorter proliferation time.

The life cycle of a cell is divided in the following stages: G0 where the cell has just emerged from mitosis and is growing to reach its mature stage; G1, the most prolonged stage where the cell does not divide and functions as part of the tissue where it belongs; S stage where the cell enters the mitotic stage and duplicates its DNA and finally the M stage where the cell enters mitosis.

Most oncogenic processes exert their greatest effect by targeting particular regulators of the G1 to S phase progression. When a cell exits from the G1 phase to enter the S phase it is

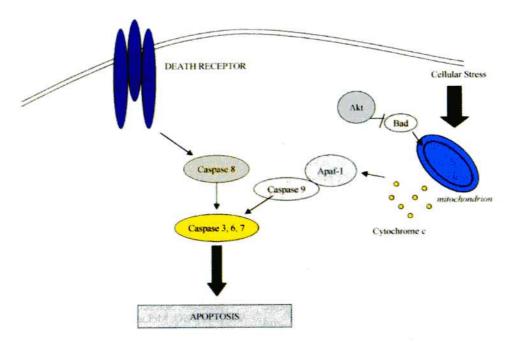


Figure 4. Regulation of apoptosis. Apoptosis can be triggered either by sensors of cellular stress or through ligation of the death receptors with their ligands. Activation of the apoptotic mechanism involves proteolytic cleavage and activation of the caspase family of proteases.

bound to divide. Control of the G1 to S progression is, therefore, a crucial checkpoint for the cell fate. Deregulation of the checkpoint proteins can contribute to uncontrolled proliferation (70). Progression from the G1 to S phase occurs when cyclins respond to growth factor signals. Thus, such signals can be initiated by different growth stimuli that transmit the signal to the cytoplasm where cyclins are bound to cyclin dependent kinases and control the restriction point. Release of the cyclin dependent kinases from the complex pinpoints the passage from G1 to S phase. Cyclins D1, D2 and D3 are known to be involved in controlling this stage. They are bound to the cyclin dependent kinases CDK4 and CDK6 which, when released, phosphorylate the retinoblastoma protein Rb (71). Phosphorylation of Rb seems to be a critical point in the cell cycle progression since it appears to be necessary for the transcriptional initiation of several genes. Hyper-phosphorylated form of Rb is present past the G1 to S restriction point and all through the cell cycle until cell division (72). The cyclin/CDK complex is inhibited by a family of proteins that include p15, p16, p18 and p19, frequently mutated in human melanomas, gliomas and leukemias, that specifically interact with CDK4 and CDK6 and therefore block the function of D type cyclins (73,74). On the other hand the p21, p27, and p57 family of cyclin inhibitors are capable of interacting with cyclins type D, E and A exhibiting a broader spectrum of inhibition (75-77) (Fig. 3).

Mutations in genes that regulate the cell cycle have been detected in several types of tumors. Inactivation of the Rb gene is a primary event in retinoblastomas (78), but overall the gene is targeted more often in adult cancers, particularly small-cell carcinomas of the lung (79). Similarly, inherited loss of INK4a gene that encodes p16 confers susceptibility to melanoma (80). Cyclin D1 is also overexpressed in many human cancers as a result of gene amplification or translocations targeting the D1

locus on human chromosome 11q13 (81). The gene encoding its catalytic partner CDK4, located on chromosome 12q13 is also amplified in sarcomas and gliomas (73) although several other potential oncogenes including MDM2, the p53 antagonist, map on the same region (82).

Although cell cycle transition depends on the underlying CDK cycle, superimposed checkpoint controls help ensure that certain processes are completed before others begin. The role of such mechanisms is to brake the cell cycle in the face of stress and damage and allowing repair to take place. The best-studied checkpoint regulator is the p53 gene and is most frequently mutated in human cancer (83,84). Even though p53 is a short-lived protein, it stabilizes and accumulates when the cell undergoes damage (85). The precise signal transduction pathway that activates p53 has not been elucidated but is likely to include genes such as ATM (mutated in ataxia telangiectasia) (86). The p53 protein acts as a transcription factor and cancer-related mutations cluster in its binding domain (85).

6. Regulation of apoptosis

Cytoplasmic or nuclear kinases transduce mitogenic signals that lead to cell proliferation and transformation. Activation of such molecules may also inhibit cells to undergo programmed cell death. Cells that suffer from DNA damage, environmental stress, or lose their ability to maintain homeostasis due to mutations, are destined to die. Apoptosis is regulated by a mechanism that involves cytochrome c release from the mitochondria and subsequent activation of several proteolytic molecules termed caspases that lead to degradation of cellular components, DNA cleavage ('laddering') and death (87). Ligation of the Fas or the TNF-α receptors with their ligands initiate signals that lead to caspase 8 activation, cyto-

chrome c release from the cytoplasm, activation of caspase 9 and the APAF complex and subsequent cleavage and activation of caspase 3, caspase 6 or caspase 7 (88,89) (Fig. 4). Caspases also translocate into the nucleus triggering their pro-apoptotic effects (89).

In cancer cells an anti-apoptotic mechanism is often activated to rescue the transformed cell from programmed cell death. The most common mechanism is activation of the bcl-2 family of proteins (Bcl-2, Bcl-xL, Bcl-W) that are able to inhibit cytochrome c release from the mitochondria and rescue the cell from apoptosis. Inactivation of the pro-apoptotic molecules Bax, Bak, Bid or Bim also contributes to rescuing the cell from apoptosis. Activation of oncogenic kinases such as Akt-1 protects cells from apoptosis by inhibiting the proapoptotic molecule Bad (50). Several anti-apoptotic signals such as growth factors (PDGF, EGF etc.) lead to the activation of signaling pathways including the PI3Kinase or MAPK pathways that can also be activated by oncogenic kinases such as Akt and Tpl-2. Thus, activation of these oncogenic kinases rescues the cell from the apoptotic signals and promotes survival.

7. Concluding remarks

The paths that a cell can take to become malignant are variable. Each different cancer type and tissue requires activation of a different set of oncogenes in order to promote transformation. These oncogenes initiate or transduce signals that lead to deregulation of gene expression and uncontrolled proliferation. Kinases are key molecules in all signaling cascades since they deliver phosphates, the 'currency' of most signaling pathways. Different kinases are being activated in different human tumors, either by being directly mutated or due to other events that induce signals of the pathway(s) they participate in. Involvement of kinases in vital signaling events renders them valuable targets for therapeutic intervention in human cancers.

References

- 1. Fearon ER: Human cancer syndromes: clues to the origin and nature of cancer. Science 278: 1043-1050, 1997.
- Hanahan D and Weinberg RA: The hallmarks of cancer. Cell 100: 57-70, 2000.
- 3. Denhardt DT: Oncogene-initiated aberrant signaling engenders the metastatic phenotype: synergistic transcription factor interactions are targets for cancer therapy. Crit Rev Oncog 7: 261-291, 1996.
- 4. Stehelin D, Varmus HE, Bishop JM and Vogt PK: DNA related to the transforming gene(s) of avian sarcoma viruses is present in normal avian DNA. Nature 260: 170-173, 1976.
- Hunter T and Cooper JA: Protein-tyrosine kinases. Annu Rev Biochem 54: 897-930, 1985.
- Kolibaba KS and Druker BJ: Protein tyrosine kinases and cancer. Biochim Biophys Acta 1333: F217-F248, 1997.
- 7. Rodrigues GA and Park M: Oncogenic activation of tyrosine kinases. Curr Opin Genet Dev 4: 15-24, 1994.
- 8. Cooper JA and Howell B: The when and how of Src regulation. Cell 73: 1051-1054, 1993.
- 9. Weiner DB, Liu J, Cohen JA, Williams WV and Greene MI: A point mutation in the neu oncogene mimics ligand induction of receptor aggregation. Nature 339: 230-231, 1989.
- Bargmann CI and Weinberg RA: Oncogenic activation of the neu-encoded receptor protein by point mutation and deletion. EMBO J 7: 2043-2052, 1988.
- 11. Zong CS, Poon B, Chen J and Wang LH: Molecular and biochemical bases for activation of the transforming potential of the proto-oncogene c-ros. J Virol 67: 6453-6462, 1993.

- 12. Sawyers CL and Denny CT: Chronic myelomonocytic leukemia: Tel-a-kinase what Ets all about. Cell 77: 171-173, 1994.
- 13. Groffen J, Stephenson JR, Heisterkamp N, de Klein A, Bartram CR and Grosveld G: Philadelphia chromosomal breakpoints are clustered within a limited region, bcr, on chromosome 22. Cell 36: 93-99, 1984.
- Heisterkamp N, Stam K, Groffen J, de Klein A and Grosveld G: Structural organization of the bcr gene and its role in the Ph' translocation. Nature 315: 758-761, 1985.
- translocation. Nature 315: 758-761, 1985.

 15. Ben-Neriah Y, Daley GQ, Mes-Masson AM, Witte ON and Baltimore D: The chronic myelogenous leukemia-specific P210 protein is the product of the bcr/abl hybrid gene. Science 233: 212-214, 1986.
- Shtivelman E, Lifshitz B, Gale RP and Canaani E: Fused transcript of abl and bcr genes in chronic myelogenous leukaemia. Nature 315: 550-554, 1985.
- 17. Hermans A, Heisterkamp N, von Linden M, van Baal S, Meijer D, van der Plas D, Wiedemann LM, Groffen J, Bootsma D and Grosveld G: Unique fusion of bcr and c-abl genes in Philadelphia chromosome positive acute lymphoblastic leukemia. Cell 51: 33-40, 1987.
- 18. Lugo TG, Pendergast AM, Muller AJ and Witte ON: Tyrosine kinase activity and transformation potency of bcr-abl oncogene products. Science 247: 1079-1082, 1990.
- Daley GQ, McLaughlin J, Witte ON and Baltimore D: The CML-specific P210 bcr/abl protein, unlike v-abl, does not transform NIH/3T3 fibroblasts. Science 237: 532-535, 1987.
 Pendergast AM, Quilliam LA, Cripe LD, Bassing CH, Dai Z, Li N, Parameters and Company and Company
- Pendergast AM, Quilliam LA, Cripe LD, Bassing CH, Dai Z, Li N, Batzer A, Rabun KM, Der CJ, Schlessinger J, et al: BCR-ABLinduced oncogenesis is mediated by direct interaction with the SH2 domain of the GRB-2 adaptor protein. Cell 75: 175-185, 1993.
- 21. Papadopoulos P, Ridge SA, Boucher CA, Stocking C and Wiedemann LM: The novel activation of ABL by fusion to an ets-related gene, TEL. Cancer Res 55: 34-38, 1995.
- ets-related gene, TEL. Cancer Res 55: 34-38, 1995.

 22. Golub TR, Goga A, Barker GF, Afar DE, McLaughlin J, Bohlander SK, Rowley JD, Witte ON and Gilliland DG: Oligomerization of the ABL tyrosine kinase by the Ets protein TEL in human leukemia. Mol Cell Biol 16: 4107-4116, 1996.
- 23. Golub TR, McLean T, Stegmaier K, Carroll M, Tomasson M and Gilliland DG: The TEL gene and human leukemia. Biochem Biophys Acta 1288: M7-M10, 1996.
- 24. Golub TR, Barker GF, Lovett M and Gilliland DG: Fusion of PDGF receptor beta to a novel ets-like gene, tel, in chronic myelomonocytic leukemia with t(5;12) chromosomal translocation. Cell 77: 307-316, 1994.
- 25. Berkowicz M, Rosner E, Rechavi G, Mamon Z, Neuman Y, Ben-Bassat I and Ramot B: Atypical chronic myelomonocytic leukemia with eosinophilia and translocation (5;12). A new association. Cancer Genet Cytogenet 51: 277-278, 1991.
- Lerza R, Castello G, Sessarego M, Cavallini D and Pannacciulli I: Myelodysplastic syndrome associated with increased bone marrow fibrosis and translocation (5;12)(q33;p12.3). Br J Haematol 82: 476-477, 1992.
- 27. Chakravarthy D, Green AR, Green VL, Kerin MJ and Speirs V: Expression and secretion of TGF-beta isoforms and expression of TGF-β-receptors I, II and III in normal and neoplastic human breast. Int J Oncol 15: 187-194, 1999.
- Kinzler KW and Vogelstein B: Lessons from hereditary colorectal cancer. Cell 87: 159-170, 1996.
- Zachos G and Spandidos DA: Expression of ras proto-oncogenes: regulation and implications in the development of human tumors. Crit Rev Oncol Hematol 26: 65-75, 1997.
- 30. Spandidos DA and Anderson ML: A role of ras oncogenes in carcinogenesis and differentiation. Adv Exp Med Biol 265: 127-131, 1990.
- 31. Okuda K, Matulonis U, Salgia R, Kanakura Y, Druker B and Griffin JD: Factor independence of human myeloid leukemia cell lines is associated with increased phosphorylation of the proto-oncogene Raf-1. Exp Hematol 22: 1111-1117, 1994.
- 32. Schmidt CA, Oettle H, Ludwig WD, Serke S, Pawlaczyk-Peter B, Wilborn F, Binder LT, Huhn D and Siegert W: Overexpression of the Raf-1 proto-oncogene in human myeloid leukemia. Leuk Res 18: 409-413, 1994.
- 33. Cheng JQ, Godwin AK, Bellacosa A, Taguchi T, Franke TF, Hamilton TC, Tsichlis PN and Testa JR: AKT2, a putative oncogene encoding a member of a subfamily of protein-serine/threonine kinases, is amplified in human ovarian carcinomas. Proc Natl Acad Sci USA 89: 9267-9271, 1992.

- Bellacosa A, de Feo D, Godwin AK, Bell DW, Cheng JQ, Altomare DA, Wan M, Dubeau L, Scambia G, Masciullo V, et al: Molecular alterations of the AKT2 oncogene in ovarian and breast carcinomas. Int J Cancer 64: 280-285, 1995.
 Ruggeri BA, Huang L, Wood M, Cheng JQ and Testa JR:
- 35. Ruggeri BA, Huang L, Wood M, Cheng JQ and Testa JR: Amplification and overexpression of the AKT2 oncogene in a subset of human pancreatic ductal adenocarcinomas. Mol Carcinog 21: 81-86, 1998.
- 36. Nakatani K, Thompson DA, Barthel A, Sakaue H, Liu W, Weigel RJ and Roth RA: Up-regulation of Akt3 in estrogen receptor-deficient breast cancers and androgen-independent prostate cancer lines. J Biol Chem 274: 21528-21532, 1999.
- Sourvinos G, Tsatsanis C and Spandidos DA: Overexpression of the Tpl-2/Cot oncogene in human breast cancer. Oncogene 18: 4968-4973, 1999.
- 38. Ohara R, Hirota S, Onoue H, Nomura S, Kitamura Y and Toyoshima K: Identification of the cells expressing cot protooncogene mRNA. J Cell Sci 108: 97-103, 1995.
- 39. Patriotis C, Makris A, Chernoff J and Tsichlis PN: Tpl-2 acts in concert with Ras and Raf-1 to activate mitogen-activated protein kinase. Proc Natl Acad Sci USA 91: 9755-9759, 1994.
- Salmeron A, Ahmad TB, Carlile GW, Pappin D, Narsimhan RP and Ley SC: Activation of MEK-1 and SEK-1 by Tpl-2 protooncoprotein, a novel MAP kinase kinase kinase. EMBO J 15: 817-826, 1996.
- 41. Tsatsanis C, Patriotis C, Bear SE and Tsichlis PN: The Tpl-2 protooncoprotein activates the nuclear factor of activated T cells and induces interleukin 2 expression in T cell lines. Proc Natl Acad Sci USA 95: 3827-3832, 1998.
- Ballester A, Tobena R, Lisbona C, Calvo V and Alemany S: Cot kinase regulation of IL-2 production in Jurkat T cells. J Immunol 159: 1613-1618, 1997.
- 43. Tsatsanis C, Patriotis C and Tsichlis PN: Tpl-2 induces IL-2 expression in T-cell lines by triggering multiple signalling pathways that activate NFAT and NF-kappaB. Oncogene 17: 2609-2618, 1998.
- 44. Lin X, Cunningham ET Jr, Mu Y, Geleziunas R and Greene WC: The proto-oncogene Cot kinase participates in CD3/CD28 induction of NF-kappaB acting through the NF-kappaB-inducing kinase and IkappaB kinases. Immunity 10: 271-280, 1999.
- Belich MP, Salmeron A, Johnston LH and Ley SC: TPL-2 kinase regulates the proteolysis of the NF-kappaB-inhibitory protein NF-kappaB1 p105. Nature 397: 363-368, 1999.
- Bellacosa A, Testa JR, Staal SP and Tsichlis PN: A retroviral oncogene, akt, encoding a serine-threonine kinase containing an SH2-like region. Science 254: 274-277, 1991.
- 47. Franke TF, Yang SI, Chan TO, Datta K, Kazlauskas A, Morrison DK, Kaplan DR and Tsichlis PN: The protein kinase encoded by the Akt proto-oncogene is a target of the PDGF-activated phosphatidylinositol 3-kinase. Cell 81: 727-736, 1995.
- 48. Chan TO, Rittenhouse SE and Tsichlis PN: AKT/PKB and other D3 phosphoinositide-regulated kinases: kinase activation by phosphoinositide-dependent phosphorylation. Annu Rev Biochem 68: 965-1014, 1999.
- 49. Bellacosa A, Chan TO, Ahmed NN, Datta K, Malstrom S, Stokoe D, McCormick F, Feng J and Tsichlis PN: Akt activation by growth factors is a multiple-step process: the role of the PH domain. Oncogene 17: 313-325, 1998.
- Khwaja A: Akt is more than just a Bad kinase. Nature 401: 33-34, 1999.
- 51. Wang HG, Pathan N, Ethell IM, Krajewski S, Yamaguchi Y, Shibasaki F, McKeon F, Bobo T, Franke TF and Reed JC: Ca²⁺-induced apoptosis through calcineurin dephosphorylation of BAD. Science 284: 339-343, 1999.
- 52. Franke TF, Kaplan DR and Cantley LC: PI3K: downstream AKTion blocks apoptosis. Cell 88: 435-437, 1997.
- 53. Ozes ON, Mayo LD, Gustin JA, Pfeffer SR, Pfeffer LM and Donner DB: NF-kappaB activation by tumour necrosis factor requires the Akt serine-threonine kinase. Nature 401: 82-85, 1999.
- 54. Romashkova JA and Makarov SS: NF-kappaB is a target of AKT in anti-apoptotic PDGF signalling. Nature 401: 86-90, 1999.
- 55. Ahmed NN, Grimes HL, Bellacosa A, Chan TO and Tsichlis PN: Transduction of interleukin-2 antiapoptotic and proliferative signals via Akt protein kinase. Proc Natl Acad Sci USA 94: 3627-3632, 1997.
- Kennedy SG, Wagner AJ, Conzen SD, Jordan J, Bellacosa A, Tsichlis PN and Hay N: The PI 3-kinase/Akt signalling pathway delivers an anti-apoptotic signal. Genes Dev 11: 701-713, 1997.

- Zimmermann S and Moelling K: Phosphorylation and regulation of Raf by Akt (protein kinase B). Science 286: 1741-1744, 1999
- 58. Beals CR, Sheridan CM, Turck CW, Gardner P and Crabtree GR: Nuclear export of NF-ATc enhanced by glycogen synthase kinase-3. Science 275: 1930-1934, 1997.
- 59. Dittmer J and Nordheim A: Ets transcription factors and human disease. Biochim Biophys Acta 1377: F1-11, 1998.
 60. Sovak MA, Bellas RE, Kim DW, Zanieski GJ, Rogers AE,
- 60. Sovak MA, Bellas ŘE, Kim DW, Zanieski GJ, Rogers AE, Traish AM and Sonenshein GE: Aberrant nuclear factor-kappaB/ Rel expression and the pathogenesis of breast cancer. J Clin Invest 100: 2952-2960, 1997.
- 61. Bukowski RM, Rayman P, Uzzo R, Bloom T, Sandstrom K, Peereboom D, Olencki T, Budd GT, McLain D, Elson P, Novick A and Finke JH: Signal transduction abnormalities in T lymphocytes from patients with advanced renal carcinoma: clinical relevance and effects of cytokine therapy. Clin Cancer Res 4: 2337-2347, 1998.
- 62. Wang W, Abbruzzese JL, Evans DB, Larry L, Cleary KR and Chiao PJ: The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. Clin Cancer Res 5: 119-127, 1999.
- 63. Schmidt EV: The role of c-myc in cellular growth control. Oncogene 18: 2988-2996, 1999.
- 64. Bouchard C, Staller P and Eilers M: Control of cell proliferation by Myc. Trends Cell Biol 8: 202-206, 1998.
- Brandt-Rauf PW and Pincus MR: Molecular markers of carcinogenesis. Pharmacol Ther 77: 135-148, 1998.
- 66. Nesbit CE, Tersak JM and Prochownik EV: MYC oncogenes and human neoplastic disease. Oncogene 18: 3004-3016, 1999.67. Spandidos DA, Karayiannis M, Yiagnisis M, Papadimitriou K and
- 67. Spandidos DA, Karayiannis M, Yiagnisis M, Papadimitriou K and Field JK: Immunohistochemical analysis of the expression of the c-myc oncoprotein in human stomach cancers. Digestion 50: 127-134, 1991.
- 68. Porter MJ, Field JK, Leung SF, Lo D, Lee JC, Spandidos DA and van Hasselt CA: The detection of the c-myc and ras oncogenes in nasopharyngeal carcinoma by immunohistochemistry. Acta Otolaryngol 114: 105-109, 1994.
- 69. Agnantis NJ, Mahera H, Maounis N and Spandidos DA: Immunohistochemical study of ras and myc oncoproteins in apocrine breast lesions with and without papillomatosis. Eur J Gynaecol Oncol 13: 309-315, 1992.
- 70. Sherr CJ: Cancer cell cycles. Science 274: 1672-1677, 1996.
- Morgan DO: Principles of CDK regulation. Nature 374: 131-134, 1995.
- 72. Zhu X, Ohtsubo M, Bohmer RM, Roberts JM and Assoian RK: Adhesion-dependent cell cycle progression linked to the expression of cyclin D1, activation of cyclin E-cdk2, and phosphorylation of the retinoblastoma protein. J Cell Biol 133: 391-403, 1996.
- 73. Nobori T, Miura K, Wu DJ, Lois A, Takabayashi K and Carson DA: Deletions of the cyclin-dependent kinase-4 inhibitor gene in multiple human cancers. Nature 368: 753-756, 1994
- gene in multiple human cancers. Nature 368: 753-756, 1994.
 74. Zhang SY, Klein-Szanto AJ, Sauter ER, Shafarenko M, Mitsunaga S, Nobori T, Carson DA, Ridge JA and Goodrow TL: Higher frequency of alterations in the p16/CDKN2 gene in squamous cell carcinoma cell lines than in primary tumors of the head and neck. Cancer Res 54: 5050-5053, 1994.
- 75. Toyoshima H and Hunter T: p27, a novel inhibitor of G1 cyclin-Cdk protein kinase activity, is related to p21. Cell 78: 67-74, 1994.
- Lee MH, Reynisdottir I and Massague J: Cloning of p57KIP2, a cyclin-dependent kinase inhibitor with unique domain structure and tissue distribution. Genes Dev 9: 639-649, 1995.
- 77. El-Deiry WS, Tokino T, Velculescu VE, Levy DB, Parsons R, Trent JM, Lin D, Mercer WE, Kinzler KW and Vogelstein B: WAF1, a potential mediator of p53 tumor suppression. Cell 75: 817-825, 1993.
- Knudson AG Jr: Mutation and cancer: statistical study of retinoblastoma. Proc Natl Acad Sci USA 68: 820-823, 1971.
- 79. Sumitomo K, Shimizu E, Shinohara A, Yokota J and Sone S: Activation of RB tumor suppressor protein and growth suppression of small cell lung carcinoma cells by reintroduction of p16INK4A gene. Int J Oncol 14: 1075-1080, 1999.
- 80. Palmero I and Peters G: Perturbation of cell cycle regulators in human cancer. Cancer Surv 27: 351-367, 1996.
- 81. Masciullo V, Scambia G, Marone M, Giannitelli C, Ferrandina G, Bellacosa A, Benedetti Panici P and Mancuso S: Altered expression of cyclin D1 and CDK4 genes in ovarian carcinomas. Int J Cancer 74: 390-395, 1997.

- 82. Hall M and Peters G: Genetic alterations of cyclins, cyclindependent kinases, and Cdk inhibitors in human cancer. Adv Cancer Res 68: 67-108, 1996.
- 83. Baker SJ, Fearon ER, Nigro JM, Hamilton SR, Preisinger AC, Jessup JM, van Tuinen P, Ledbetter DH, Barker DF, Nakamura Y, et al: Chromosome 17 deletions and p53 gene mutations in colorectal carcinomas. Science 244: 217-221, 1989.
- colorectal carcinomas. Science 244: 217-221, 1989.

 84. Nigro JM, Baker SJ, Preisinger AC, Jessup JM, Hostetter R, Cleary K, Bigner SH, Davidson N, Baylin S, Devilee P, et al: Mutations in the p53 gene occur in diverse human tumour types. Nature 342: 705-708, 1989.
- Ko LJ and Prives C: p53: puzzle and paradigm. Genes Dev 10: 1054-1072, 1996.
- 86. Enoch T and Norbury C: Cellular responses to DNA damage: cell-cycle checkpoints, apoptosis and the roles of p53 and ATM. Trends Biochem Sci 20: 426-430, 1995.
- 87. Green DR: Apoptotic pathways: the roads to ruin. Cell 94: 695-698, 1998.
- 88. Qin H, Srinivasula SM, Wu G, Fernandes-Alnemri T, Alnemri ES and Shi Y: Structural basis of procaspase-9 recruitment by the apoptotic protease-activating factor 1. Nature 399: 549-557, 1999.
- 89. Alnemri ES: Hidden powers of the mitochondria. Nature Cell Biol 1: E40-E42, 1999.