

RESEARCH ARTICLE

Yin Yang-1(YY-1) expression in idiopathic pulmonary fibrosis

Giorgos A Margaritopoulos^{1,2}, Katerina M Antoniou^{1,2}, Giannoula Soufla³, Evi Vassalou², Demetrios A Spandidos³, and Nikos M Siafakas¹

¹Interstitial Lung Disease Unit, Department of Thoracic Medicine, University Hospital of Heraklion, Crete, Greece, ²Laboratory of Molecular and Cellular Pulmonary Medicine, Medical School, University of Crete, Heraklion, Crete, Greece, and ³Department of Virology, Medical School, University of Crete, Heraklion, Crete, Greece

Abstract

Context: Yin Yang-1 (YY-1) is implicated in the pathogenesis of lung cancer which can be complicated with idiopathic pulmonary fibrosis (IPF).

Objective: The aim of the study was to investigate whether YY-1 is involved in the pathogenesis of IPF and whether represents a common pathogenetic pathway which could explain the coexistence of these disorders.

Materials and methods: Lung tissue from 52 patients (37 with IPF and 15 controls) and bronchoalveolar lavage fluid (BALF) from 34 patients (25 with IPF and 9 controls) were studied and YY-1 mRNA expression was evaluated by realtime PCR.

Results: YY-1 was expressed in 8% (3/37) of IPF patients and in 6% (1/15) of healthy controls in tissue samples. In addition, 12% (3/25) of IPF patients and 33% (3/9) of healthy controls have expressed YY-1 gene in BALF samples. However, no statistical significant difference in mRNA expression between patients and controls has been detected in both tissue and BAL fluid samples.

Discussion and conclusion: Our results do not support the hypothesis of YY-1 involvement in IPF. However, similar expression of YY-1 gene in two biological samples cannot exclude a possible role of this polymorphic gene in the pathway of IPF. Further studies in a larger scale of patients are needed.

Keywords: Yin Yang-1, transcription factor, tumorigenesis, idiopathic pulmonary fibrosis, lung cancer

Introduction

Yin Yang-1 (YY-1) is a ubiquitous zinc-finger transcription factor that has the ability to initiate as well as to regulate the transcription through activation or repression (1). It has an important role in various biological processes such as cell-cycle control, embryogenesis, apoptosis, B-cell development and tumorigenesis (1).

A growing body of evidence has underlined the role of YY-1 in various steps during the process of tumorigenesis. YY-1 interacts with cell-cycle regulation via its association with the tumor suppressor gene p53 (2,3) and activates, when overexpressed, the protooncogenes c-myc which are implicated in the pathogenesis of various cancers (4,5). It also negatively regulates Fas transcription and expression and confers resistance to Fas-induced apoptosis (6) and moreover is implicated in tumor progression and metastasis (7,8). In addition, it might be responsible for the resistance of various cancers to chemotherapeutic agents (9).

Several studies in various types of lung cancer have shown that YY-1 is upregulated (10-12) suggesting a possible role of this factor in the pathogenesis of lung cancer. Idiopathic pulmonary fibrosis (IPF), another elusive lung disease with fatal outcome, can be complicated with lung cancer (13-15) raising questions regarding the similarity of IPF and lung cancer biology. Surprisingly, some pathogenetic hallmarks such as genetic alterations, uncontrolled and aberrant cellular proliferation and tissue invasion are common in both diseases leading to the hypothesis that IPF might be considered as a neoproliferative disorder of the lung (16).

Address for Correspondence: Katerina M Antoniou, MD, PhD, Lecturer of Thoracic Medicine, Department of Thoracic Medicine, University Hospital, Medical School, University of Crete, Heraklion, 71110 Crete, Greece. Tel.: +30-2810 392 433; Fax: +30 2810 542 650. E-mail: katerinaantoniou@yahoo.gr



However, the YY-1 gene expression has not been explored in IPF. The purpose of our study was to shed light on the role of YY-1 transcription factor in the pathogenesis of this lethal disorder.

Patients and methods

Patients

We studied the lung tissue from 52 patients (37 with IPF and 15 controls, prospectively collected in our institute from 2000 to 2010) and the bronchoalveolar lavage fluid (BALF) from 34 patients (25 with IPF and 9 controls, prospectively collected from 2006 to 2010).

Tissue specimens and BALF were obtained from consenting individuals in accordance with institutional review board approval. Patients with IPF had clinical and radiographic findings consistent with the diagnosis of IPF, or pathologic confirmation of the diagnosis of usual interstitial pneumonia. None of the patients in either the IPF or control group had been previously treated or were currently being treated with corticosteroids or other immunosuppressive agents. The control lung tissue specimens were from subjects undergoing thoracic surgery for reasons other than interstitial lung disease, mainly for lung cancer. The lung tissue was obtained from a site as distant as possible from the primary tumor as previously described (17) and was histologically free of neoplasm.

RNA extraction and reverse transcription

Total RNA was extracted from each lung specimen and from BAL fluid using a power homogenizer and the TRIzol® reagent (Invitrogen, Carlsband, CA) according to the manufacturer's instructions. cDNA was synthesized using the Strascript reverse transcriptase kit (Stratagene, La Jolla, CA) as previously described(18).

Real-time RT-PCR

YY-1 mRNA expression was measured using a real-time RT-PCR assay with SYBR-Green I. Beta-αctin (β-actin) was used as the internal control, in order to normalize YY-1 expression levels, as previously described (18). Primer sequences and their annealing temperatures are described in Table 1. For all the tested samples, the expression of both the target and the housekeeping gene was calculated, projecting, with the help of the standard curve, the Ct value of all unknown samples to a relative mRNA quantity. The mRNA gene expression of every sample was normalized by dividing the mRNA value of the target gene with the appropriate β -actin mRNA value according to the following formula:

$$\frac{\text{Normalized Sample}}{\text{control}} = \frac{(1 + E_{gene})^{-\Delta C}{}_{t}Gene}{(1 + E_{Actin})^{-\Delta C}{}_{t}Actin}$$

Statistical analysis

YY-1 mRNA levels were first evaluated by the one-sample Kolmogorov-Smirnov goodness of fit test, in order to determine whether they follow a normal distribution pattern. The t-test was used to compare the mRNA expression of YY-1 after stratification for normal, lung and IPF samples. The Chi-square (χ^2) test was used to evaluate the significant statistical differences in YY-1 mRNA expression status. All statistical analyses were performed with SPSS 11.5 (SPSS, Chicago, IL). Statistical significance was set at the 95% level (P < 0.05).

Results

In tissue samples, YY-1 was expressed in 8% (3/37) of IPF patients and in 6% (1/15) of healthy controls (Table 2).

In BAL fluid samples, YY-1 was expressed in 12% (3/25) of IPF patients and in 33% (3/9) of healthy controls

We have not detected any statistical significant difference in mRNA expression between the study's group in both tissue and BAL fluid samples.

Discussion

The aim of our study was to investigate the role of YY-1 transcription factor in the pathogenesis of IPF and, since it is well known that this factor is implicated in the process of tumorigenesis, to seek indirect evidence that its activity could explain the coexistence of IPF and lung cancer.

To the best of our knowledge, this is the first attempt to evaluate the expression of YY-1 in lung tissue and BAL fluid of patients with IPF. We have observed that YY-1 was expressed in a small number of patients and controls. Additionally, no statistical significant difference in mRNA expression has been found between patient's group and controls in both types of samples. However, a similar rather low expression of the YY-1 gene has been detected in both biological samples, lung tissue and BAL fluid of patients with IPF. These results do not support our hypothesis but can not also exclude the involvement of YY-1 in the pathogenesis of IPF.

Table 1. Primer sequences used for quantitative real-time RT-PCR

Gene	Primer pair Sequence (5′–3′)	Annealing temperature
YY-1	For: GGAATACCTGGCATTGACC	58°C
	Rev: TCTTTGTGCAGCCTTTGAG	
β-Actin	For: CGGCATCGTCACCAACTG	58°C
	Rev: GGCACACGCAGCTCATTG	

Table 2. YY-1 expression in IPF and controls in lung tissue and BAL.

	IPF	CONTROLS	P value
YY-1 (tissue)	3/37 (8%)	1/15 (6%)	NS
YY-1 (BAL)	3/25 (12%)	3/9 (33%)	NS

 χ^2 test: P < 0.05 is considered statistically significant. NS, not significant.



The role of YY-1 in cancer biology is contradictory because this factor can act as a transcriptional activator or repressor depending on the context in which it binds (1). YY-1 overexpression is associated with an increased malignant phenotype and poor outcome in breast cancer, cervical neoplasia, osteosarcoma, myeloid leukaemia and Hodgkin's and non-Hodgkin's lymphoma (19). On the contrary, higher YY-1 protein levels were associated with a favorable prognosis in prostate and ovarian cancers(19).

Regarding lung cancer, gene expression analysis of YY-1 has revealed an increase of mRNA levels in squamous cell carcinoma and in adenocarcinoma in comparison to normal tissue (10-12). Unpublished data from our laboratory have shown that 4/17 (23.5%) of the patients with lung cancer have expressed YY-1 in BAL fluid with no significant difference in mRNA expression between patients' group and controls.

Recently, our study group investigated the role of epidermal growth factor receptor (EGFR) and its downstream signalling pathways including the phosphoinositide 3-kinase (PI3K)/Akt/mammalian target of rapamycin (mTOR) and Ras/Raf/MEK/ERK (mitogenactivated protein kinase (MAPK) pathway in the pathogenesis of IPF (18,20). EGFR as well as the Akt and MAPK pathways have a key role during tumorigenesis (21-24). We found no significant different in mRNA expression of EGFR between IPF and healthy subjects, whereas a downregulation of K-ras and an upregulation of B-raf has been observed in IPF patients in comparison to controls suggesting that despite the weak expression, these oncogenes could be implicated in the pathogenesis of IPF.

A growing body of evidence has pointed out several pathways common in these two disorders. Mutations of p53 gene, a tumor suppressor gene altered in most cases of cancer, have been demonstrated also in IPF (25). Mutations in the components of telomerase, an enzyme aimed to maintain telomere length, as well as telomere shortening, events so far almost exclusive of tumor development, have been described in both familial and sporadic IPF (26-28). The oxidant/antioxidant balance is implicated in the pathogenesis of IPF. Oxidative stress which activates the synthesis of several profibrotic cytokines leading to tissue injury and fibrosis, may cause alterations in DNA methylation patterns and specific histone modifications triggering the process of tumorigenesis (29). Intracellular signaling pathways crucial in the pathogenesis of lung cancer and mesothelioma (30) such as Wnt/bcatenin are fully involved in IPF (31).

Conclusion

Our findings do not support but can not also exclude the hypothesis that YY-1 gene is involved in the pathogenesis of IPF, representing a common pathogenetic pathway between IPF and lung cancer. The link between cancer and IPF is becoming intriguing and may give birth to new approaches regarding the pathogenesis and therapy of IPF. For years, it was believed that fibrogenesis was linked to a single pathogenetic pathway, and this belief led to the failure of trials targeting an individual pathway (32,33). The hypothesis of multiple pathogenetic pathways has been adopted in cancer biology, and targeting simultaneously these pathways is the main thinking in cancer therapy (34-36). Should this hypothesis be adopted in order to shed light in the pathogenesis and design therapeutic trials for fibrotic lung disorders and in particular for IPF(37)? Further investigations are needed to verify this hypothesis.

Declaration of interest

The authors report no declarations of interest.

References

- Zaravinos A, Spandidos DA. Yin yang 1 expression in human tumors. Cell Cycle 2010, 9, 512-522
- Yakovleva T, Kolesnikova L, Vukojevic V, Gileva I, Tan-No K, Austen M, Lüscher B, Ekström TJ, Terenius L, Bakalkin G. YY1 binding to a subset of p53 DNA-target sites regulates p53-dependent transcription. Biochem Biophys Res Commun 2004, 318, 615-624.
- Sui G, Affar el B, Shi Y, Brignone C, Wall NR, Yin P, Donohoe M, Luke MP, Calvo D, Grossman SR, Shi Y. Yin Yang 1 is a negative regulator of p53. Cell 2004, 117, 859-872.
- Riggs KJ, Saleque S, Wong KK, Merrell KT, Lee JS, Shi Y, Calame K. Yin-yang 1 activates the c-myc promoter. Mol Cell Biol 1993, 13, 7487-7495
- 5. Shrivastava A, Yu J, Artandi S, Calame K. YY1 and c-Myc associate in vivo in a manner that depends on c-Myc levels. Proc Natl Acad Sci USA 1996, 93, 10638-10641.
- Garbán HJ, Bonavida B. Nitric oxide inhibits the transcription repressor Yin-Yang 1 binding activity at the silencer region of the Fas promoter: a pivotal role for nitric oxide in the up-regulation of Fas gene expression in human tumor cells. J Immunol 2001, 167,
- 7. Brankin B, Skaar TC, Brotzman M, Trock B, Clarke R. Autoantibodies to the nuclear phosphoprotein nucleophosmin in breast cancer patients, Cancer Epidemiol Biomarkers Prev 1998, 7, 1109-1115.
- Beier UH, Görögh T, Implications of galactocerebrosidase and galactosylcerebroside metabolism in cancer cells. Int J Cancer 2005, 115, 6-10.
- 9. Baritaki S, Huerta-Yepez S, Sakai T, Spandidos DA, Bonavida B. Chemotherapeutic drugs sensitize cancer cells to TRAIL-mediated apoptosis: up-regulation of DR5 and inhibition of Yin Yang 1. Mol Cancer Ther 2007, 6, 1387-1399.
- 10. Wachi S, Yoneda K, Wu R. Interactome-transcriptome analysis reveals the high centrality of genes differentially expressed in lung cancer tissues. Bioinformatics 2005, 21, 4205-4208.
- 11. Takeuchi T, Tomida S, Yatabe Y, Kosaka T, Osada H, Yanagisawa K et al. Expression profile-defined classification of lung adenocarcinoma shows close relationship with underlying major genetic changes and clinicopathologic behaviors. J Clin Oncol. 2006, 10, 1679-1688,
- 12. Jones MH, Virtanen C, Honjoh D, Miyoshi T, Satoh Y, Okumura S et al. Two prognostically significant subtypes of high-grade lung neuroendocrine tumours independent of small-cell and large-cell $neuro endocrine\ carcino mas\ identified\ by\ gene\ expression\ profiles.$ Lancet 2004, 6, 775-781
- 13. Turner-Warwick M, Lebowitz M, Burrows B, Johnson A. Cryptogenic fibrosing alveolitis and lung cancer. Thorax 1980, 35, 496-499

- 14. Matsushita H, Tanaka S, Saiki Y, Hara M, Nakata K, Tanimura S, Banba J. Lung cancer associated with usual interstitial pneumonia. Pathol Int 1995, 45, 925-932.
- 15. Hubbard R, Venn A, Lewis S, Britton J. Lung cancer and cryptogenic fibrosing alveolitis. A population-based cohort study. Am J Respir Crit Care Med 2000, 161, 5-8.
- 16. Vancheri C, Failla M, Crimi N, Raghu G. Idiopathic pulmonary fibrosis: a disease with similarities and links to cancer biology. Eur Respir J 2010, 35, 496-504.
- 17. Mehrad B, Burdick MD, Zisman DA, Keane MP, Belperio JA, Strieter RM. Circulating peripheral blood fibrocytes in human fibrotic interstitial lung disease. Biochem Biophys Res Commun 2007, 353, 104-108,
- 18. Antoniou KM, Soufla G, Lymbouridou R, Economidou F, Lasithiotaki I, Manousakis M, Drositis I, Spandidos DA, Siafakas NM. Expression analysis of angiogenic growth factors and biological axis CXCL12/CXCR4 axis in idiopathic pulmonary fibrosis. Connect Tissue Res 2010, 51, 71-80.
- 19. Castellano G, Torrisi E, Ligresti G, Malaponte G, Militello L, Russo AE, McCubrey JA, Canevari S, Libra M. The involvement of the transcription factor Yin Yang 1 in cancer development and progression. Cell Cycle 2009, 8, 1367-1372.
- 20. Antoniou KM, Margaritopoulos GA, Soufla G, Symvoulakis E, Vassalou E, Lymbouridou R, Samara KD, Kappou D, Spandidos DA, Siafakas NM, Expression analysis of Akt and MAPK signaling pathways in lung tissue of patients with idiopathic pulmonary fibrosis (IPF). J Recept Signal Transduct Res 2010, 30, 262-269.
- 21. Weihua Z, Tsan R, Huang WC, Wu Q, Chiu CH, Fidler IJ, Hung MC. Survival of cancer cells is maintained by EGFR independent of its kinase activity. Cancer Cell 2008, 13, 385-393.
- 22. Sos ML, Fischer S, Ullrich R, Peifer M, Heuckmann JM, Koker M, et al. Identifying genotype-dependent efficacy of single and combined PI3K and MAPK-pathway inhibition in cancer. Proc Natl Acad Sci USA 2009, 106, 18351-18356.
- 23. Engelman JA, Chen L, Tan X, Crosby K, Guimaraes AR, Upadhyay R, Maira M, McNamara K, Perera SA, Song Y, Chirieac LR, Kaur R, Lightbown A, Simendinger J, Li T, Padera RF, García-Echeverría C, Weissleder R, Mahmood U, Cantley LC, Wong KK. Effective use of PI3K and MEK inhibitors to treat mutant Kras G12D and PIK3CA H1047R murine lung cancers. Nat Med 2008, 14, 1351-1356.
- 24. Thannickal VJ, Horowitz JC. Evolving concepts of apoptosis in idiopathic pulmonary fibrosis. Proc Am Thorac Soc 2006, 3, 350-356.

- 25. Kuwano K, Kunitake R, Kawasaki M, Nomoto Y, Hagimoto N, Nakanishi Y, Hara N. P21Waf1/Cip1/Sdi1 and p53 expression in association with DNA strand breaks in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 1996, 154, 477-483.
- 26. Cronkhite JT, Xing C, Raghu G, Chin KM, Torres F, Rosenblatt RL, Garcia CK. Telomere shortening in familial and sporadic pulmonary fibrosis. Am J Respir Crit Care Med 2008, 178, 729-737.
- 27. Liu T, Chung MJ, Ullenbruch M, Yu H, Jin H, Hu B, Choi YY, Ishikawa F, Phan SH. Telomerase activity is required for bleomycin-induced pulmonary fibrosis in mice. J Clin Invest 2007, 117, 3800-3809.
- 28. Tsakiri KD, Cronkhite JT, Kuan PJ, Xing C, Raghu G, Weissler JC, Rosenblatt RL, Shay JW, Garcia CK. Adult-onset pulmonary fibrosis caused by mutations in telomerase. Proc Natl Acad Sci USA 2007, 104, 7552-7557.
- 29. Franco R, Schoneveld O, Georgakilas AG, Panayiotidis MI. Oxidative stress, DNA methylation and carcinogenesis. Cancer Lett 2008, 266, 6-11.
- 30. Mazieres J, He B, You L, Xu Z, Jablons DM. Wnt signaling in lung cancer. Cancer Lett 2005, 222, 1-10.
- 31. Königshoff M, Balsara N, Pfaff EM, Kramer M, Chrobak I, Seeger W, Eickelberg O. Functional Wnt signaling is increased in idiopathic pulmonary fibrosis. PLoS ONE 2008, 3, e2142.
- 32. Raghu G, Brown KK, Bradford WZ, Starko K, Noble PW, Schwartz DA, King TE Jr; Idiopathic Pulmonary Fibrosis Study Group. A placebo-controlled trial of interferon gamma-1b in patients with idiopathic pulmonary fibrosis. N Engl J Med 2004, 350, 125-133.
- 33. King TE, Behr J, Brown KK, du Bois RM, Raghu G. Bosentan use in idiopathic pulmonary fibrosis (IPF): results of the placebo controlled BUILD-1 study. Am J Respir Crit Care Med 2006, 3, A524.
- 34. Benito M, Diaz-Rubio E. Molecular biology in colorectal cancer. Clin Transl Oncol 2006, 8, 391-398.
- 35. Colebatch A, Hitchins M, Williams R, Meagher A, Hawkins NJ, Ward RL. The role of MYH and microsatellite instability in the development of sporadic colorectal cancer. Br J Cancer 2006, 95,
- 36. Pao W, Wang TY, Riely GJ, Miller VA, Pan Q, Ladanyi M, Zakowski MF, Heelan RT, Kris MG, Varmus HE. KRAS mutations and primary resistance of lung adenocarcinomas to gefitinib or erlotinib. PLoS Med 2005, 2, e17.
- 37. Maher TM, Wells AU, Laurent GJ. Idiopathic pulmonary fibrosis: multiple causes and multiple mechanisms? Eur Respir J 2007, 30, 835-839

